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Range Characterization Studies at Donnelly Training Area, Alaska: 2001 and 2002

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Front cover: The 4/11 Field Artillery preparing to fire an M119A 105-mm howitzer.

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ABSTRACT

The U.S. Army Alaska seeks to conserve and protect natural resources on lands used for combat training exercises. Some of these exercises require live fire of ordnance containing high explosives. One aspect of managing the ranges so as to mitigate the environmental consequences of training is to identify the location, extent, and potential migration of munitions residues in soils, surface waters, and groundwater. This report summarizes analytical results for soil samples collected from firing points and some impact areas at the Donnelly Training Area near Delta Junction, Alaska. Explosives residues are for the most part undetectable or at very low concentrations (parts per billion) in the soils of impact areas. The exceptions are soils near or under partial ordnance detonations, targets, and rocket motor debris. We found high concentrations (parts per thousand) of TNT in soils next to partially detonated 500-lb and 2000-lb bombs; moderate concentrations (parts per million) of RDX and TNT around targets; and moderate concentrations (parts per million) of NG under rocket motor debris. At firing points used for 105-mm howitzers, 2,4-DNT is detectable in surface soils at parts-per-million concentrations. This analyte is associated with burned and unburned fibers of propellant that are sprayed to distances of at least 100 m from the muzzle. The highest concentrations of 2,4-DNT were in soils where excess propellant is burned for disposal. Because of the very low soil clean-up levels listed by the State of Alaska for this compound, appropriate and reproducible laboratory and field sampling procedures need to be developed to monitor this analyte.

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PREFACE

This report was prepared by Marianne E. Walsh, Chemical Engineer, Environmental Sciences Branch, Cold Regions Research and Engineering Laboratory (CRREL), Engineer Research and Development Center (ERDC); Charles M. Collins, Research Physical Scientist, Environmental Sciences Branch, CRREL; Alan D. Hewitt, Research Physical Scientist, Environmental Sciences Branch, CRREL; Michael R. Walsh, Mechanical Engineer, Engineering Resources Branch, CRREL; Thomas F. Jenkins, Research Chemist, Environmental Sciences Branch, CRREL; Jeffrey Stark, formerly Physical Science Technician, Civil and Infrastructure Engineering Branch, CRREL; Arthur Gelvin, Engineering Technician, Engineering Resources Branch, CRREL; Thomas A. Douglas, Research Chemist, Environmental Sciences Branch, CRREL; Nancy Perron, Physical Science Technician, Snow and Ice Branch, CRREL; Dennis Lambert, Mechanical Engineering Technician, Engineering Resources Branch, CRREL; Ronald Bailey, Biological Sciences Technician, Environmental Sciences Branch, CRREL; and Karen Myers, Biologist, Environmental Laboratory.

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1 INTRODUCTION

The withdrawal of training lands from the public domain on Fort Wainwright and Donnelly Training Area (formerly Fort Greely) in Interior Alaska was renewed under the Military Lands Withdrawal Act (PL106-65). As part of the Environmental Impact Statement prepared for the renewal, the Army pledged to assess the amount of residues from explosive munitions at the currently used testing and training impact ranges in Donnelly Training Area and Fort Wainwright and the potential for surface water and groundwater contamination (U.S. Army Alaska 1999). The training lands of Fort Greely were renamed the Donnelly Training Area in 2001 when Fort Greely was realigned under the Base Realignment and Closure (BRAC) process. The main post area of Fort Greely was slated for closure, while the training lands were transferred administratively to Fort Wainwright. Subsequently, the Fort Greely main post has been withdrawn from BRAC and transferred to the Army Space and Missile Defense Command to support the Ground-Based Mid-Course Intercept Missile Defense (GMD) Program. Donnelly Training Area has 26,300 hectares (or 263 km²) of impact areas where high-explosive ammunition is used, including the Washington and Mississippi Impact Areas located within the floodplain of the Delta River, the Delta Creek Impact Area located within the floodplain of Delta Creek, and the Oklahoma Impact Area located just east of Delta Creek.

Assessing the levels of explosives residues by sampling the soil and water is a challenge because of the large size and varied terrain of these impact areas, the safety hazards associated with unexploded ordnance, and on-going live-fire training. Of most interest is the potential for contamination of surface water and groundwater that would provide a route for migration of the explosives residues

across military installation boundaries. We used an authoritative sampling strategy (sample locations were selected based on prior knowledge) to identify explosives source areas within the impact areas. In our opinion, authoritative sampling is a more efficient approach to the overall goal of protecting water sources than random sampling, which is used when there is little or no information about the potential distribution of the analytes of interest.

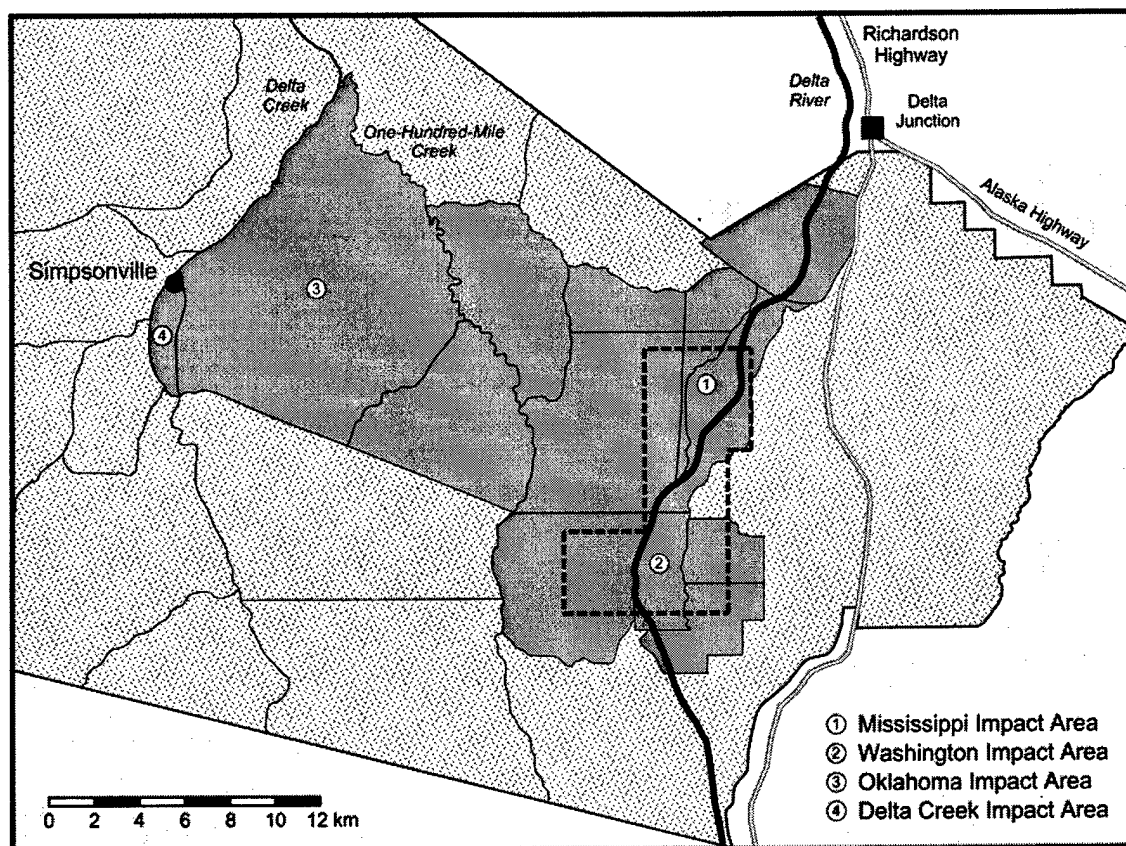
During July 2000, we undertook the initial sampling program on Washington Impact Area and Lampkin Range (Walsh et al. 2001), where we selected, based on guidance from the Cold Regions Test Center, specific locations within the impact area where known ordnance items had detonated. We collected discrete and multi-increment samples to determine if we could find any explosives residues in the surface soils. We detected explosives residues in 48% of the samples we collected, most frequently RDX and TNT. Concentrations were low (the median concentrations for RDX and TNT were 21 and 5 $\mu\text{g/kg}$, respectively) except where ordnance items failed to detonate completely and solid chunks of explosives were on the surface soil. We also detected propellant residues (2,4-DNT and NG) at the Lampkin Range firing point.

2 OBJECTIVES

In 2001, the objective of the sampling was to determine if we could detect any explosives residues and source areas that could contribute to groundwater contamination in the Donnelly Training Area. The impact areas that we sampled were Delta Creek, Georgia Island, and Washington Range West. We also sampled several firing points to determine concentrations of propellant residues. Based on the analytical results for the 2001 firing point samples, which showed that we needed to expand our sampled collection to distances greater than 50 m from the 105-mm gun firing platforms, we collected additional firing point samples in 2002. Our objective was to characterize the distribution of propellant residues around a firing position and to monitor the persistence of the residues after 30 days of weathering. An additional objective was to obtain more depth samples to determine the potential for downward migration of the residues. Because persistence and migration are influenced by the soil matrix, we chose two firing positions for intensive sampling, one that was vegetated and one that was sparsely vegetated.

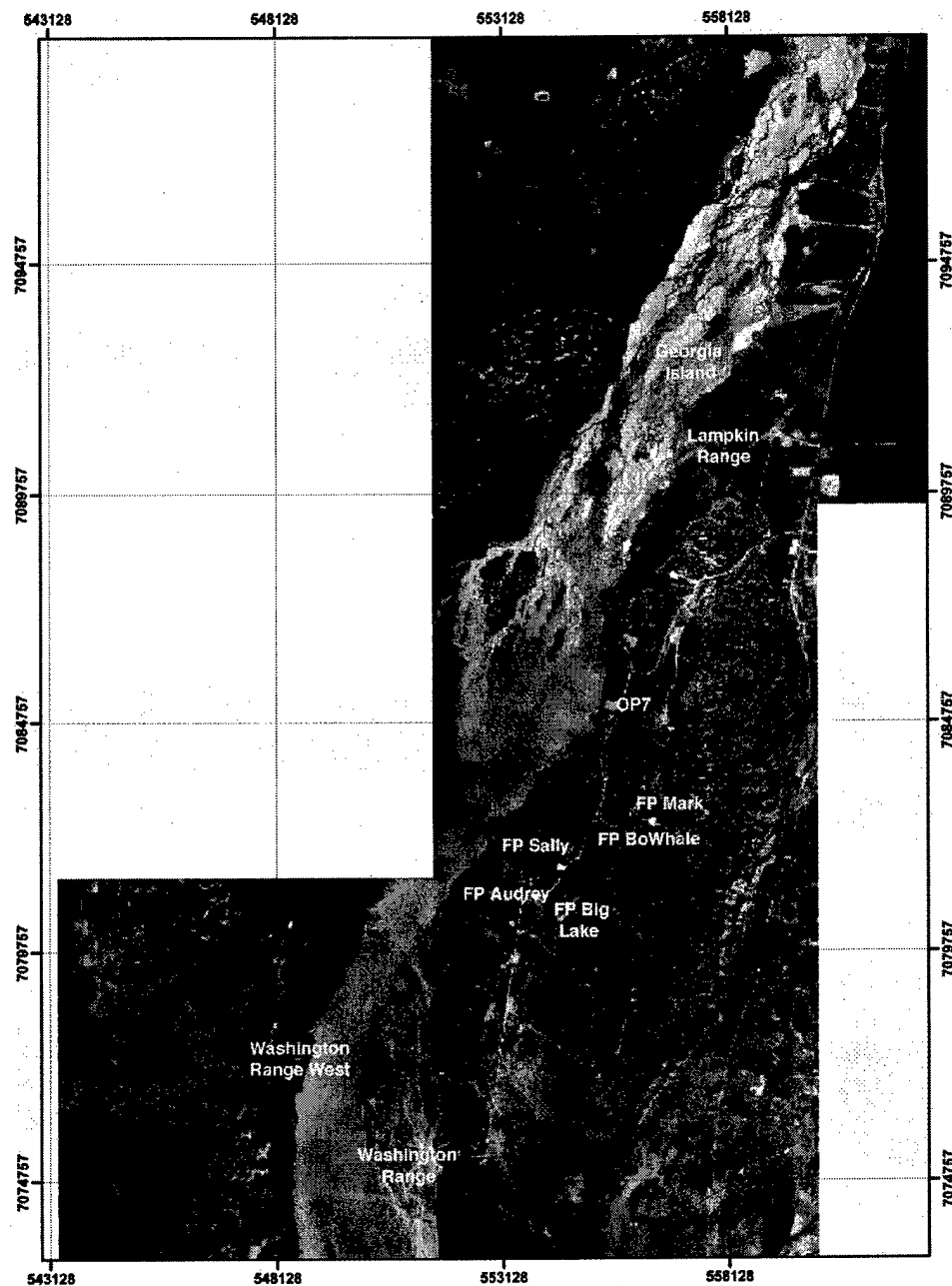
3 PHYSICAL SETTING

The Donnelly Training Area (Fig. 1) consists of 2,554 km² located in the northern foothills of the Alaska Range and the Tanana-Kuskokwim Lowlands. Several glacial outwash rivers, including the Delta River, Delta Creek, and the Little Delta River, flow northward from the Alaska Range across the training area to the Tanana River (U.S. Army Alaska 2003). Several large impact areas, totaling 263 km², are located within the training area, including the Washington and Mississippi Impact Areas along the Delta River, Oklahoma Impact Range east of Delta Creek, and Delta Creek Impact Area along Delta Creek. The Army uses Washington and Mississippi Impact Areas mainly for indirect-fire weapons (the target cannot be seen by the gunner), while Delta Creek (Table 1) and Oklahoma Impact Areas are used primarily for aerial bombing by the Air Force (U.S. Army Alaska 2002).



a. Donnelly Training Area, showing the impact areas sampled. The dashed lines indicate the area shown in Figure 1b.

Figure 1. Installation maps and orthophotos.



b. Orthophoto (AeroMap U.S. 2003), taken August 2002, showing the Delta River, the locations of firing points, Washington Range, Lampkin Range, and Georgia Island.

Figure 1 (cont.).

Table 1. Ordnance used by the Army at the impact areas and firing points that we sampled (based on 1998 to 1999 ammo reports).

| Ordnance (DODIC) | Target analyte potentially in residue | | Location used and sampled |
|--|---------------------------------------|------------------------------|---|
| | Explosive | Propellant | |
| 5.56-mm cartridges (A059, A064, A066, A075) | | NG PETN in pellet booster | FP: Simpsonville, Lampkin IA: Delta Creek |
| 7.62-mm cartridges (A107, A127) | | NG | FP: Simpsonville, Lampkin IA: Delta Creek |
| .50 caliber cartridges (A520, A555) | | NG, 2,4-DNT, PETN | FP: Simpsonville, Lampkin IA: Delta Creek |
| 30-mm cartridges (B103) | | | FP: Lampkin |
| 40-mm cartridge (B470) | RDX | NG | FP: Simpsonville, Lampkin IA: Delta Creek |
| 40-mm cartridge [B519(TP) B576 (TP) B535 (ILL), M918 (TP)] | | NG | Simpsonville, Delta Creek, Lampkin |
| 105-mm cartridges (C445) | TNT/RDX | 2,4-DNT | FP: Mark, Sally, Audrey, Bo-Whale, Lampkin, Simpsonville IA: Delta Creek |
| 105-mm cartridges [C508 (HEAT)] | TNT/RDX | NG | FP: Mark |
| 105-mm cartridges (C511) | | NG | FP: Audrey, Bo-Whale, Mark |
| 105-mm cartridges (C520) | | 2,4-DNT | FP: Mark, Bo-Whale |
| 105-mm cartridges [C449 (ILL)] | | 2,4-DNT | FP: Mark, Sally, Audrey, Bo-Whale IA: Delta Creek |
| 60-mm (B642) | TNT/RDX | NG | FP: Lampkin, OP7, Simpsonville IA: Delta Creek |
| 60-mm [B640 (ILL)] | | | FP: Lampkin, OP7, Simpsonville IA: Delta Creek |
| 81-mm [C226 (ILL)] | | NG | FP: Lampkin, OP7, Simpsonville |
| 81-mm (C256) | TNT/RDX | NG | FP: Simpsonville |
| M67 (G881) | TNT/RDX | | FP: Lampkin |
| 2.75-inch rocket [H180 (ILL)] | | NG | FP: Simpsonville IA: Delta Creek |
| Claymore mine (K143) | RDX | | FP: Lampkin, Simpsonville IA: Delta Creek |
| 84mm AT4 (C995) | M136? | | FP: Lampkin, Simpsonville IA: Delta Creek |
| 155-mm HC and ILL (D445, D505) | | | FP: Mark, Sally, Bo-Whale |
| C4 (M023) | RDX | | Lampkin, Simpsonville |
| Bangalore torpedo (M028) | RDX/TNT | | Lampkin, Simpsonville, Delta Creek |
| Detonation cord (MD15) | PETN | | Simpsonville |
| TOW (PB25) | HMX | | FP: Simpsonville IA: Delta Creek |
| Dragon (PL23) | | | FP: Simpsonville, Lampkin IA: Delta Creek |

TP: Target practice rounds that do not contain high-explosive filler.

ILL: Illumination round.

IA: Impact Area. The Mississippi and Oklahoma Impact Areas were extensively used but were not sampled due to UXO hazards.

The Delta River is a large, glacially fed, braided river that starts out as a clear-water stream draining the Tangle Lakes on the south side of the Alaska Range. It cuts across the crest of the Alaska Range, receiving meltwater from a number of glaciers, including the Canwell, Castner, and Black Rapids Glaciers. In the vicinity of Donnelly Training Area, the river cuts through the Donnelly Moraine, a late-Pleistocene moraine marking the last major glacial advance down the Delta River valley (Péwé and Holmes 1964, Péwé 1975). The incised moraine forms large bluffs on either side of the river valley. The river through this area is braided and has a broad, gravel floodplain. In the vicinity of the Washington and Mississippi Impact Areas, there are large abandoned floodplain terraces, several meters above the present active floodplain. These terraces represent episodes of greater sedimentation in the past, probably associated with surges of the Black Rapids Glacier over the last several hundred years. Much of the terrace of the Washington Range is bare gravel, with localized areas of sparse shrubs mostly consisting of silverberry (*Eleagnus commutata*). Jorgenson et al. (2001) mapped the vegetation on Fort Greely and classified these areas as riverine gravelly barrens and riverine gravelly low scrub and dry dwarf scrub.

Delta Creek is also a glacially fed braided river that flows from the Alaska Range north, joining the Tanana River. It receives meltwater from the Trident and Hayes Glacier, as well as snowmelt from the Alaska Range. Like the Delta River, it has extensive sections of abandoned floodplain terraces several meters higher than the current active braided floodplain. One-Hundred-Mile Creek is a small, single-channel, clear-water stream originating in the foothills of the Alaska Range and flowing northward and then westward, joining Delta Creek. The Delta Creek Impact Area (Fig. 2), a 20-km² impact area, is located along 9 km of Delta Creek. Target arrays are located along abandoned floodplain terraces on the west side of the active creek. The western boundary of Oklahoma Impact Area, a 250-km² impact area, is located along 16 km of Delta Creek, north of Delta Creek Impact Range. The eastern and northern boundary of Oklahoma Impact Area runs along One-Hundred-Mile Creek. Simpsonville (Fig. 3) is a Military Operations in Urban Terrain/Combined Arms Live Fire Exercise (MOUT/CALFEX) site located on top of a bluff on the west bank of Delta Creek. The gently sloping area is mostly open, covered with tussock tundra vegetation.

The western side of Washington Impact Area is along the west bank of the Delta River. Here a narrow floodplain runs along the steep bluffs of the moraine to the west. The narrow floodplain is vegetated with lowland gravelly dry mixed forest (Jorgenson et al. 2001) and shows little evidence of artillery use, such as cratering or range scrap, probably because of its location at the edge of the impact area. Georgia Island (Fig. 4) is a 4-km-long island within the active floodplain of the Delta River. It is sparsely to heavily vegetated [classified as riverine gravelly barrens to lowland gravelly dry mixed forest by Jorgenson et al.

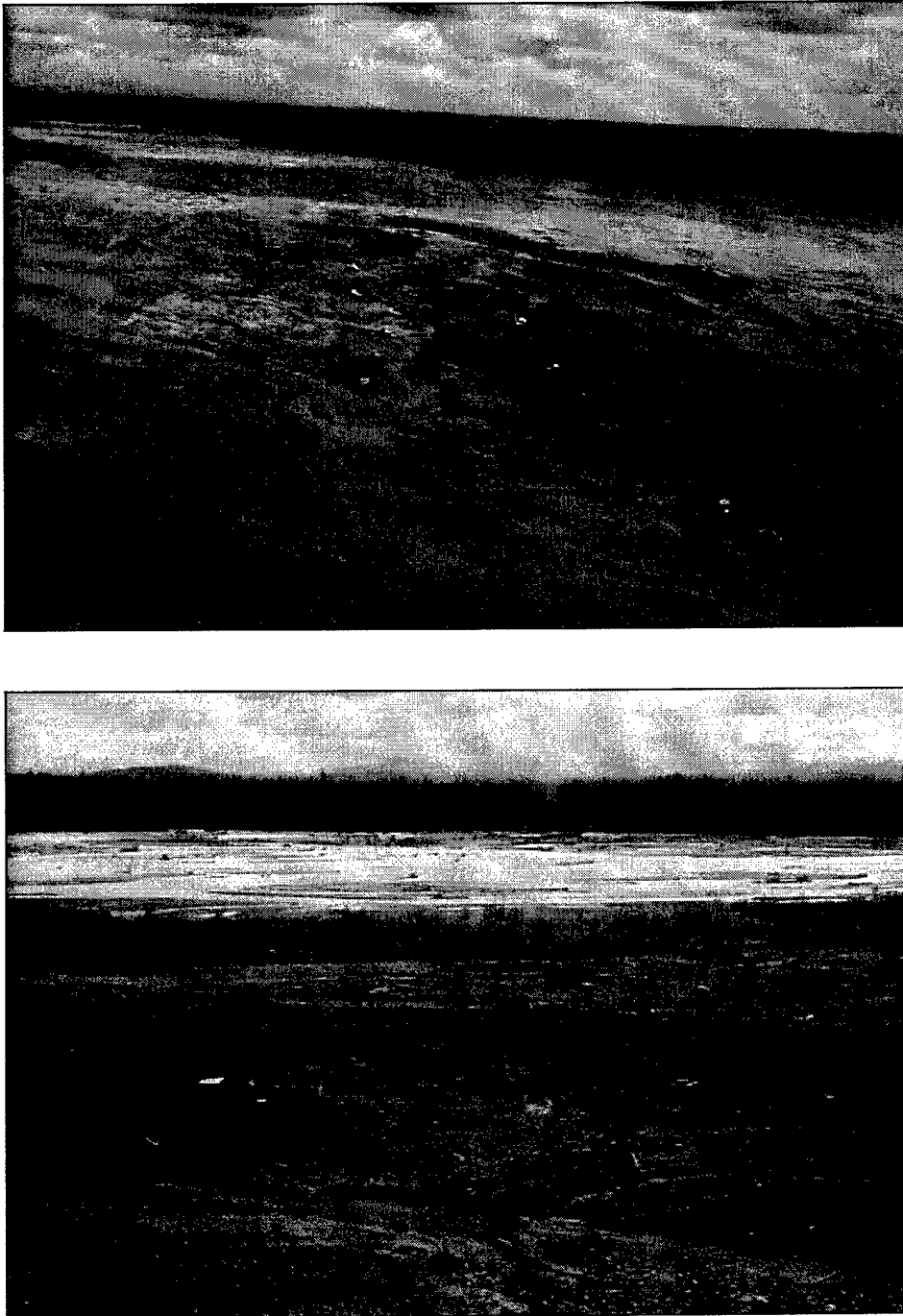


Figure 2. Aerial and near-ground views of a target array located 2 km downstream of Delta Creek Impact Area.



Figure 3. Aerial view of Simpsonville MOUT/CALFEX, located on a bluff overlooking the Delta Creek Impact Area.



Figure 4. Aerial view of Georgia Island, showing the old target berm.

(2001)]. It is located immediately downstream of Mississippi Impact Area, a heavily used indirect fire range where we are not allowed to sample because of extreme UXO (unexploded ordnance) hazards. Georgia Island has been used to a lesser degree as an artillery impact area. It has also been used as a target area for direct-fire weapons from various ranges on the east side of the Delta River.

Firing Points Audrey, Bo-Whale, Big Lake, Mark, and Sally are located in the Donnelly East Training Area on the east side of the Delta River (Fig. 1b). The firing points are located on either side of Meadows Road, which runs south along the broad crest of the glacial lateral moraine forming the high bluffs on the east side of the river. The firing points are used for indirect fire into the Mississippi and Washington Impact Areas to the west. FP Big Lake, Bo-Whale, and Sally (Fig. 5a) are open vegetated areas with a ground cover of grasses, sedges, low forbs, and some low shrubs. Soils are fine-grained silt loam overlying coarser, poorly sorted gravel. The soils at FP Bo-Whale are wetter and have more organic material than those of the other firing points. FP Mark (Fig. 5b) and Audrey are mostly unvegetated open area with sporadic ground cover of mosses and grasses. Soils here are poorly sorted silty, sandy gravel. The Lampkin Range firing point (Fig. 6) is located on an elevated, broad, flat-topped gravel berm or platform built on the vegetated floodplain along the east bank of the Delta River. The berm where we sampled was constructed of silty, sandy gravel.



a. FP Sally (vegetated site), July 2002.

Figure 5. Firing points used for indirect fire into Mississippi and Washington Impact Areas.



b. FP Mark (sparsely vegetated site), July 2002.

Figure 5 (cont.).

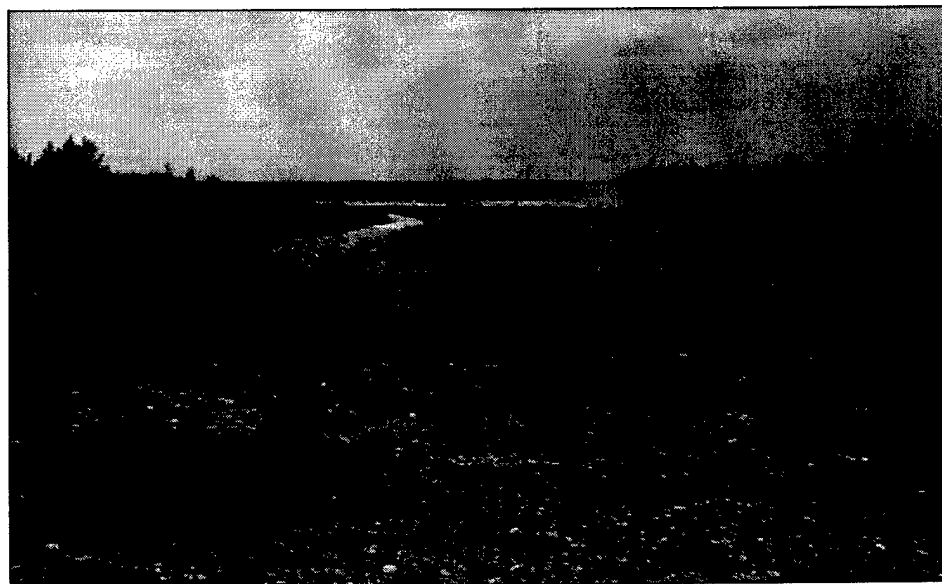


Figure 6. Ground view from Lampkin Range Firing Point, which is used for direct fire at targets within the floodplain of the Delta River.

4 METHODS

Field Sample Collection

Delta Creek, 2001

In June 2001, we collected samples downstream of the boundaries of the Delta Creek Impact Area. We were not allowed to sample the actual Delta Creek Impact Area because of the hazards associated with unexploded submunitions. However, a series of targets and associated craters and range scrap (Fig. 2) were located 2 km downstream, where we collected both discrete and composite samples. The discrete samples were soil near what appeared to be partial detonations of 500-lb bombs. The composite samples consisted of fifty 40-g subsamples collected around craters of various dimensions, around targets, and in undisturbed areas. At 5, 8, 11, 14, and 17 km downstream were suitable helicopter-landing sites with fine-grain sediments, where we collected more samples. With the exception of two discrete samples collected under pieces of rocket motors, samples farther downstream were composites from 10- × 10-m areas on inactive and abandoned bar surfaces along the edge of the creek.

We also collected seven samples at the MOUT/CALFEX site known as Simpsonville located on a bluff overlooking Delta Creek (Fig. 3). Four of the samples were from explosive ordnance disposal craters, and the other three were from craters thought to be produced by 40-mm grenades.

Georgia Island, 2001

The sampling of Georgia Island, within the Delta River, was conducted by sampling approximately every 200 m along the centerline of the island and every 50 m along the base of a former target berm (Fig. 4). At each sampling location, a multi-increment sample was collected by taking approximately fifty 40-g random discrete subsamples over a 10- × 10-m area as was done at Delta Creek. A total of 44 composite samples were collected. Five discrete samples were collected near ordnance items such as empty 40-mm grenade casings and range scrap.

West side of Washington Impact Area, 2001

The sampling of the west side of Washington Impact Area, along the west bank of the Delta River, was to be conducted like the sampling of Georgia Island at every 200 m along the narrow vegetated floodplain. However, heavy vegetation and lack of suitable helicopter landing spots limited where we could sample

along the bank. At several locations we collected samples at 50- to 100-m intervals, walking to several sites from a single landing site. At each sampling location a sample was collected by taking approximately fifty 40-g random discrete subsamples over a 10- × 10-m area as was done at Delta Creek and Georgia Island. Twenty-four composite samples were collected.

Firing Points, 2001

Previous sampling at Fort Greely, Fort Lewis, Yakima Training Center, and other training areas has shown that firing points are frequently contaminated with propellant residues (Walsh et al. 2001). The most common residues detected have been 2,4-DNT, which is an additive in single-base propellants, and NG, an ingredient in double- and triple-base propellants (U.S. Army 1984).

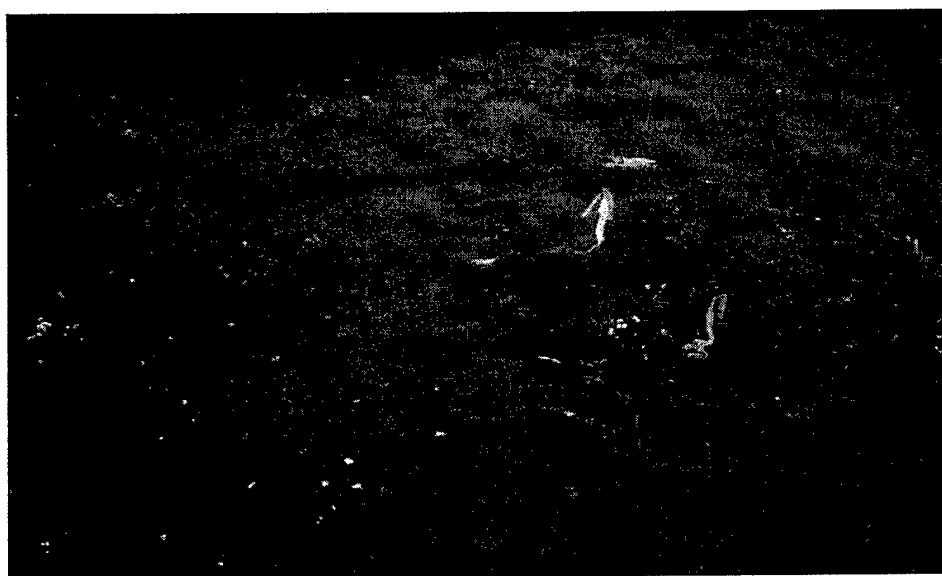
Our objective in sampling the firing points at Donnelly Training Area was to determine the average concentrations of propellant residues in the surface soil. Depending on the locations of the firing points, these residues could contaminate groundwater or be ingested by grazing animals. However, the samples we collect can be used to compute mean concentrations only if the concentration estimates for replicate samples agree within reasonable limits. Previous sampling efforts on firing ranges have indicated that concentration estimates in replicate samples can vary by more than a factor of ten. Recently, the problem of laboratory subsampling of unvegetated explosives-contaminated soil was solved by grinding soils using a ring mill, a practice routinely used in the mining industry but not in environmental laboratories. However, the problem of reproducible field sample collection has yet to be resolved.

During the week of July 31 to August 5, 2001, we sampled Donnelly East Training Area firing points that had been used during the second week of June 2001 by the 4/11 Field Artillery. About 100 rounds had been fired from M119A 105-mm howitzers at each of firing points Audrey, Sally, Big Lake, Bo-Whale, and Mark (Fig. 1). Major S. Houston accompanied us to various firing points, and he located the firing positions of several 105-mm howitzers at firing points Sally, Bo-Whale, and Big Lake. The firing positions were identified by the characteristic depressions left on the ground by the firing platform and spade of each howitzer (Fig. 7).

We collected surface samples in front of eight howitzer firing positions. First we staked a line representing the axis of the cannon tube position and parallel lines 3 m on either side (Fig. 8). At 3.5, 7, 14, 21, and 28 m distance from the center of each firing platform depression, we collected duplicate multi-increment samples. Each sample consisted of 30 increments of the surface soil and associated vegetation collected within a 1- × 6-m area. At three howitzer firing positions we collected five additional samples 50 m from the firing platform



a. M119A1 105-mm howitzers.



b. Depressions made by the firing platform and spade.

Figure 7. Locating howitzer firing positions in July 2001. The firing platform is located between the wheels and the spade is to the rear of the gun.

depression. One of these samples was along the axis of the cannon tube, and the other samples were $\pm 30^\circ$ and $\pm 60^\circ$ from the axis.

Each sample was returned to our field laboratory and air-dried on an aluminum pie pan. While the sample was drying, a subsample was taken for the field analysis described below. This analysis allowed us to identify which firing points

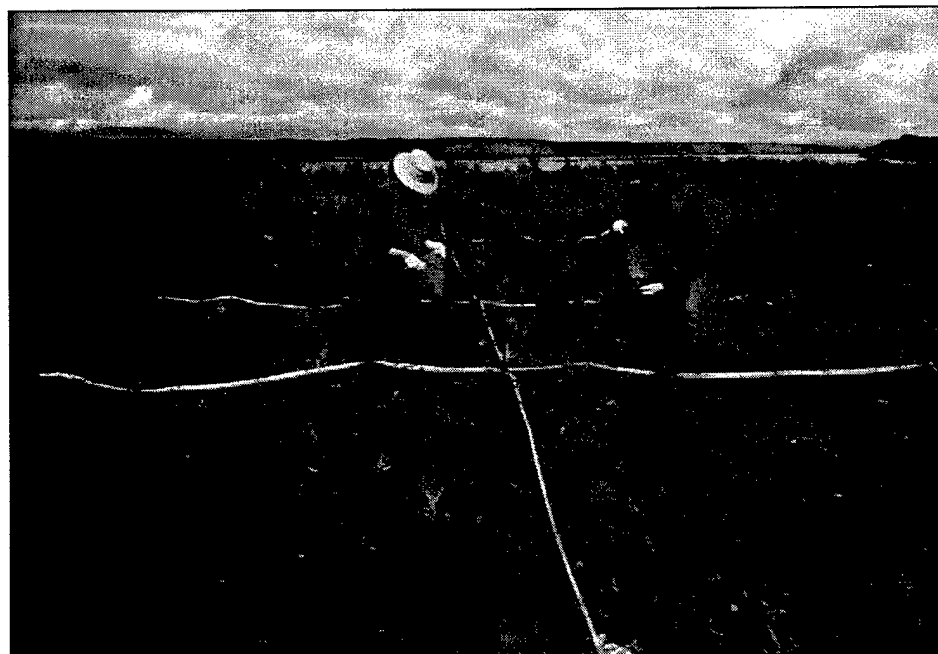


Figure 8. FP Sally in July 2001. The axis of the cannon tube corresponds to the yellow tape measure down the center of the photo. Multi-increment samples were collected within a 1- × 6-m area at 3.5, 7, 14, and 28 m from the center of the depression left by the firing platform.

had detectable concentrations of propellant residues. Based on these analyses, we returned to the sites of the samples with the four highest propellant residue concentrations and collected discrete samples and subsurface samples. Results from the field analysis also allowed us to select samples to send to CRREL (Hanover, NH) to test sample homogenization techniques. The remainder of the samples were sent to the ERDC's Environmental Lab (Vicksburg, Mississippi).

Firing Points, 2002

From June 19 to June 25, 2002, the 4/11th Field Artillery set up at the same firing points as in 2001 for indirect fire training and at the Lampkin Range for direct fire training. A. Gelvin and T. Douglas were on location for some of the firing and obtained exact howitzer positions from CPT Mandelloni of B Company. Gelvin and Douglas then started collecting six composite samples from each gun location. Each sample was nominally made up of 30 increments randomly collected with a bulb planter (Fig. 9) to a depth of 1 cm taken over a 2- × 6-m area. The sample locations were 25 and 50 m in front of each gun and at 60° left and 60° right (Fig. 10). These samples were returned to our field lab for drying, sieving, field-grinding (Hewitt and Walsh 2003), and field gas chroma-



a. Using a bulb planter.



b. Sample increment, nominally 1 cm thick.

Figure 9. Collecting surface samples at firing point Sally.

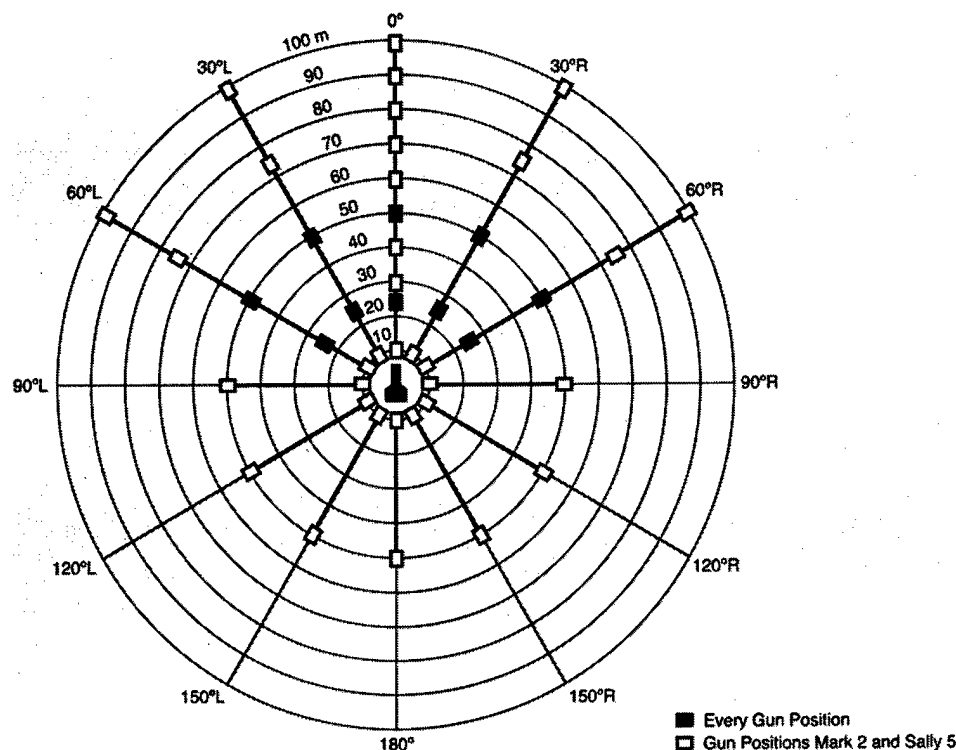


Figure 10. Sampling scheme used for characterization of propellant residues around a howitzer firing position.

topographic analysis. Based on these analyses, we chose two gun positions for intensive sampling. These positions were FP Sally Gun 5 (Fig. 5a), which was heavily vegetated, and FP Mark Gun 2 (Fig. 5b), which was sparsely vegetated. We collected samples radially every 30° at 10 and 50 m, where possible, from the gun platform location (Fig. 10). In some cases the boundary of the firing point was less than 50 m from the gun platform, so the samples were collected at the boundary. Additional samples were collected at 25-m intervals out to 100 m, where possible, $\pm 30^\circ$ and $\pm 60^\circ$ from the axis of the gun tube. Samples were collected at 10-m intervals directly in front of the gun platform.

In July 2002, we repeated the intensive sampling at FP Mark Gun 2 and FP Sally Gun 5. We also collected subsurface composite samples 25 and 50 m in front of the gun and at 60° left and 60° right. Each subsurface composite sample was made up of five increments collected at a depth of 15–20 cm using a Series 400 AMS corer.

Two additional sampling locations were OP7 (Fig. 1b), where excess propellant was burned, and the Lampkin Range firing point, where direct-fire exercises with howitzers, mortars, 40-mm grenades, and other ordnance occur (Table 1, Fig. 1b, 6).

Lab Processing of Samples

Firing Points, 2001 and 2002

Most of the firing points are located on well-vegetated fields, so the surface samples were a mix of soil, decayed organic matter, and vegetation. This very complicated matrix presented a considerable subsampling challenge. Most of the firing point samples were shipped to the ERDC Environmental Lab (Vicksburg, MS), where they were analyzed using standard homogenization methods (i.e., manual grinding with a mortar and pestle and sieving through a #30 mesh sieve). The remaining samples, which we selected based on the results of the field gas chromatographic analyses, were sent to CRREL to examine the subsampling heterogeneity associated with these surface samples and test homogenization techniques (Walsh et al. 2002). The selected samples were from a Bo-Whale firing point (Fig. 11).

First, we separated each sample into two size fractions using #10 mesh (2-mm) sieves. The <2-mm fraction consisted of soil and organic matter. The >2-mm fraction contained leafy and woody vegetation and some pebbles. We took duplicate 10-g subsamples from each size fraction of each sample for determination of propellant residues. Then we machine-ground (Fig. 12) each of the size fractions and took a second set of duplicate 10-g subsamples. The grinding, which was done for 60 s on a LabtechEssa LM2 ring mill at CRREL, reduced the particle size of the samples to less than 0.1 mm. Two of the ground samples were divided using a LabtechEssa RSD005 rotary divider.

All of the firing point samples in 2002 were sieved through a #10 (2-mm) mesh sieve, and the <2-mm fraction was machine-ground on a LabtechEssa LM2 ring mill. The grind time for vegetated samples was increased to 90 s. Duplicate 10-g subsamples were taken for analysis for each sample.

Delta Creek

All samples from Delta Creek were air-dried prior to shipment to CRREL for analysis. Those samples that were expected to contain explosives were subsampled by taking larger than normal (50-g) soil aliquots in an effort to reduce subsampling error. All others were subsampled by taking 10-g soil aliquots. The soils were extracted using acetone, and the extracts were analyzed using the colorimetric Method 8515 (U.S. EPA) to detect TNT and other nitroaromatics. This procedure was performed because some of the samples were collected near what appeared to be partial detonations of 500-lb bombs that contained TNT. We used the results of the colorimetric method to sort the samples by TNT concentration. Samples that were positive by the colorimetric method were analyzed by HPLC (see below), and all others were analyzed by GC- μ ECD. Selected samples



Figure 11. Firing position at Bo-Whale from which samples were collected for homogenization studies.

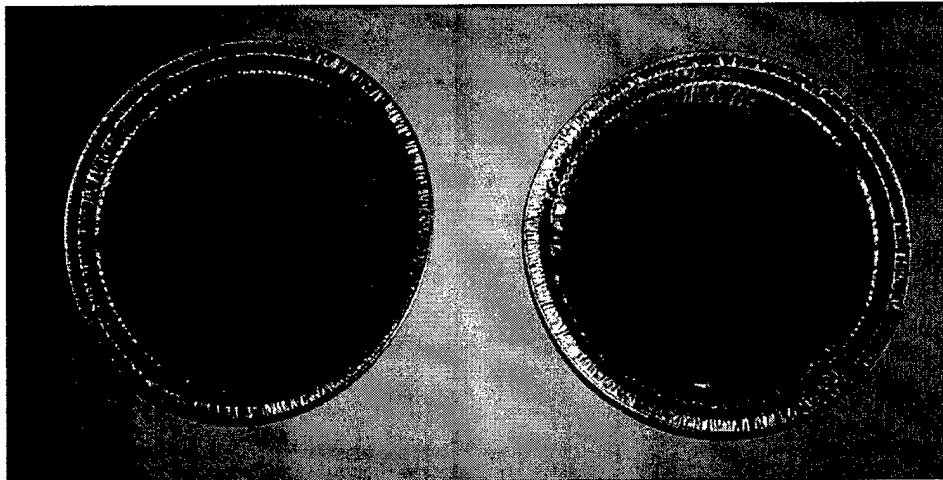


Figure 12. Unground (left) and ground (right) >2-mm fractions of a Bo-Whale sample.

(TNT concentrations between 1 and 200 $\mu\text{g}/\text{kg}$) were machine-ground on a LabTectonics ring mill at Mineral Stats, Inc. (Broomfield, Colorado) and re-analyzed for explosives. This further processing was done to reduce the subsampling error associated with explosives-contaminated soils (Walsh et al. 2002).

Analytical Methods Used by CRREL

In the field lab during the July–August 2001 and June 2002 sampling periods, acetone extracts were analyzed on a field-portable gas chromatograph equipped with a thermionic ionization detector (Hewitt et al. 2001, USEPA 2001). The SRI Model 8610C gas chromatograph has a heated injection port, and chromatographic separations were achieved on a 15-m \times 0.53-mm 100% dimethylpolysiloxane column. This procedure provides detection limits of 10 $\mu\text{g/kg}$ for TNT and 2,4-DNT and 100 $\mu\text{g/kg}$ for RDX.

In the laboratory, we used Method 8095 (Nitroaromatics and Nitramines by GC) (USEPA 2000), which uses an electron capture detector and provides detection limits near 1 $\mu\text{g/kg}$ for TNT and RDX. We used an HP 6890 and a Restek 6-m \times 0.53-mm id RTX-5ms (95% dimethyl–5% diphenyl polysiloxane) column. The method detection limits for Method 8095 are 1 $\mu\text{g/kg}$ for the di- and trinitroaromatics, 3 $\mu\text{g/kg}$ for RDX, 25 $\mu\text{g/kg}$ for HMX, 10 $\mu\text{g/kg}$ for NG, and 20 $\mu\text{g/kg}$ for PETN.

We used Method 8330 [Nitroaromatics and Nitramines by High Performance Liquid Chromatography (HPLC)] (USEPA 1994) when we found higher-concentration samples ($>0.2 \mu\text{g/g}$). The HPLC separations were achieved on a 15-cm \times 3.9-mm (4- μm) Nova Pak C₈ (Waters Millipore) column eluted with 1.4 mL/min 15:85 isopropanol:water and on a 25-cm \times 4.6-mm (5- μm) Supelco LC-CN column eluted with 1.2-mL/min 65:14:21 water:methanol:acetonitrile. Detection was by UV (254 nm).

Collection of Propellant Residue from a Snow-covered Firing Point

To further examine the deposition of propellant residues from 105-mm howitzers, we had the opportunity to collect samples in conjunction with a research project that involves detonations of ordnance items on clean snow surfaces where the snow acts as a pristine collection surface for the post-blast residues (Hewitt et al. 2003). In March 2002, seventy-one 105-mm projectiles were fired from Firing Point Neiber (Fig. 13) at Fort Richardson, AK. The propellant residues were visible on the snow surface as either fibrous black soot (Fig. 14) or unburned yellow fibers. Samples of the residues were collected by shoveling into plastic bags the top layer of snow from 1-m² areas within and just beyond the visible plume forward and to the sides of the gun muzzle. Snow samples were also collected at the breaches of three guns, where the expended cartridges are removed from the howitzer. The snow was melted, and then the particulate residue fraction was obtained by filtration through glass fiber filters. The filtrate and the solid residue were analyzed separately for 2,4-DNT.



Figure 13. Winter firing of an M119A1 105-mm howitzer.

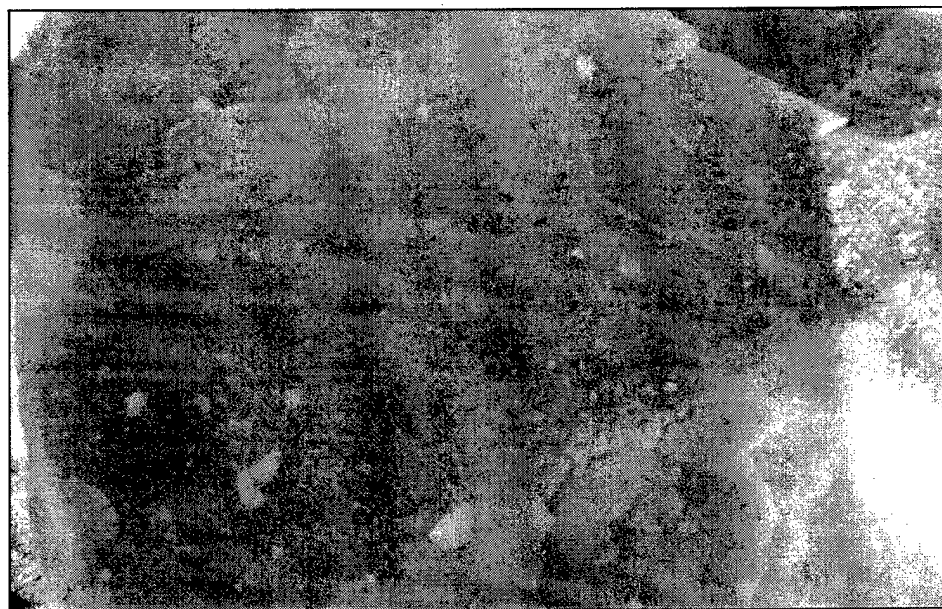


Figure 14. Fibrous residue deposited on the snow surface from the firing of a 105-mm howitzer.

5 RESULTS

Delta Creek Impact Area

Explosives residues were detected in all of the samples collected near the target array located 2 km downstream from the Delta Creek Impact Area. In the composite samples, the following residues were determined: TNT (<1–314,000 µg/kg); RDX (7–1,400 µg/kg); HMX (<25–110 µg/kg); 2,4-DNT (1–33 µg/kg), and NG (<15–51 µg/kg). Only four of the samples had TNT above 1,000 µg/kg, and the median concentration was 80 µg/kg. The amino-DNT reduction products were detected in each sample as well, but concentrations were low (<200 µg/kg). One of the discrete samples collected near a 500-lb bomb partial detonation had a TNT concentration of 17,300,000 µg/kg, a concentration far exceeding any other sample we collected. No explosives residues were detected upstream of the target array, and NG was the only propellant residue detected downstream of the target array. The NG (2,000 and 80 µg/kg) was found in two discrete samples that were collected under pieces of rocket motors.

Explosives residues were detected in each of the seven soil samples from Simpsonville, the MOUT/CALFEX site. The concentration ranges were: TNT (<d–140 µg/kg), RDX (<d–26 µg/kg), 2,4-DNT (<d–28 µg/kg), and NG (<d–1,500 µg/kg). The NG was associated with 40-mm grenade training, and the other residues were associated with explosive ordnance disposal craters.

Georgia Island

All composite samples collected along the centerline of Georgia Island and from the base of the target berm were negative for HMX, RDX, TNT, 2,4-DNT, and other target analytes. NG was detected in a discrete soil sample, GI003, taken under an empty 40-mm grenade cartridge casing. The concentration was 4,700 µg/kg.

West Side of Washington Impact Area

Explosives residues were not detectable in any of the samples from the narrow vegetated floodplain along the west side of Washington Impact Area.

Firing Points 2001

Each of the firing points that we sampled in 2001 at Donnelly Training Area had detectable concentrations of 2,4-DNT in at least one composite sample (Appendix Table 1). A typical chromatogram is shown in Figure 15. The spatial

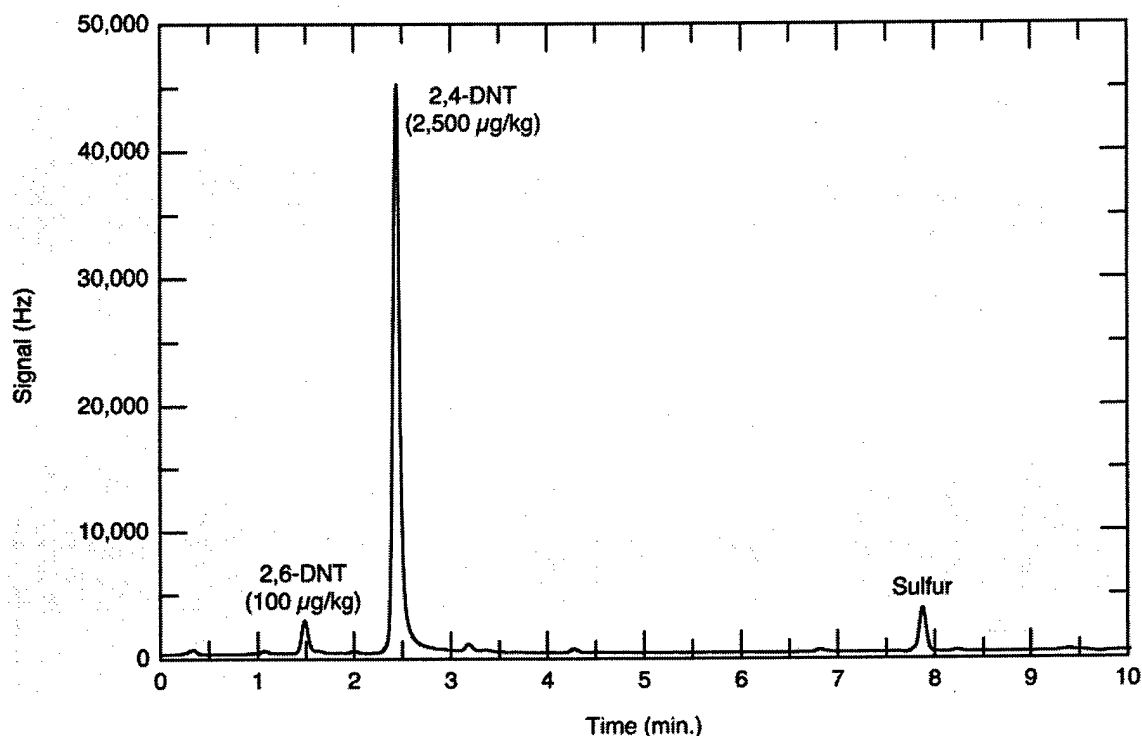


Figure 15. Typical chromatogram obtained by GC- μ ECD of an extract of a soil collected from a 105-mm howitzer firing point.

distribution of 2,4-DNT was extremely heterogeneous, as shown by the concentration estimates in discrete samples. For example, five discrete samples collected within the 1- \times 6-m area from which Bo-Whale composite sample 1 was collected ranged in concentration from 25 to 7,900 μ g/kg. There was also generally poor agreement between duplicate field samples that were processed by standard methods at EL.

Our sample homogenization experiments were done on the duplicate field samples that we collected at the Bo-Whale firing point (Fig. 11). First we took duplicate laboratory subsamples of the <2-mm and >2-mm size fractions. The >2-mm fraction is not routinely analyzed for contaminant concentrations (Paetz and Cröbmann 1994). However, the propellant residues fall onto whatever substrate is near the howitzer, so we did not feel justified in excluding any part of the surface samples we collected. We then machine-ground each size fraction to a fine powder (Fig. 12) and took duplicate subsamples for analysis.

Concentration estimates of 2,4-DNT in the machine-ground and not-ground samples are shown in Table 2. To determine if machine grinding increased subsampling precision of the two size fractions, we used an F test. First, we computed the pooled variances for the laboratory duplicates using the following equation:

Table 2. Concentrations of 2,4-DNT in laboratory subsamples of the >2-mm and <2-mm fractions with and without machine grinding. Samples were collected July 2001 from FP Bo-Whale.

| Distance from firing platform (m) | Angle from centerline (degrees) | Sample ID | Field rep. | Lab rep. | 2,4-DNT concentration (µg/kg) | | | |
|--|---------------------------------------|--------------|---------------|-------------|-------------------------------|-----------|-------------|------------|
| | | | | | Machine ground | | Not ground | |
| | | | | | >2 mm | <2 mm | >2 mm | <2 mm |
| 3.5 | 0 | 1 | A | 1 | 903 | 8,540 | 14,400 | 5,000 |
| 3.5 | 0 | 1 | A | 2 | 1,560 | 5,470 | 1,570 | 1,720 |
| 3.5 | 0 | 1 | B | 1 | 301 | 3,400 | 219 | 1,120 |
| 3.5 | 0 | 1 | B | 2 | 397 | 3,640 | 3,320 | 1,500 |
| 7 | 0 | 2 | A | 1 | 130 | 1,860 | 369 | 1,700 |
| 7 | 0 | 2 | A | 2 | 143 | 2,550 | 1,070 | 3,800 |
| 7 | 0 | 2 | B | 1 | 1,270 | 3,030 | 3,230 | 6,500 |
| 7 | 0 | 2 | B | 2 | 623 | 3,660 | 131 | 972 |
| 14 | 0 | 3 | A | 1 | 483 | 1,750 | 299 | 580 |
| 14 | 0 | 3 | A | 2 | 616 | 732 | 136 | 157 |
| 14 | 0 | 3 | B | 1 | 84 | 1,400 | 68 | 2,470 |
| 14 | 0 | 3 | B | 2 | 224 | 2,000 | 123,000 | 11,600 |
| 21 | 0 | 4 | A | 1 | 450 | 1,280 | <d | 96 |
| 21 | 0 | 4 | A | 2 | 485 | 1,120 | <d | 984 |
| 21 | 0 | 4 | B | 1 | 2,400 | 1,520 | 440 | 36 |
| 21 | 0 | 4 | B | 2 | 1,940 | 2,300 | 140 | 356 |
| 28 | 0 | 5 | A | 1 | 3,870 | 16,900 | 12,900 | 29,000 |
| 28 | 0 | 5 | A | 2 | 3,450 | 29,900 | 9,430 | 16,500 |
| 28 | 0 | 5 | B | 1 | 10,800 | 24,000 | 11,100 | 12,500 |
| 28 | 0 | 5 | B | 2 | 15,300 | 29,100 | 9,450 | 6,300 |
| 50 | -30 | 6 | | 1 | 172 | 4,020 | 14 | 5,980 |
| 50 | -30 | 6 | | 2 | 193 | 2,840 | 104 | 2,030 |
| 50 | -15 | 7 | | 1 | 200 | 8,320 | 477 | 2,310 |
| 50 | -15 | 7 | | 2 | 186 | 5,860 | 843 | 2,630 |
| 50 | 0 | 8 | | 1 | 4,510 | 6,790 | 1,670 | 794 |
| 50 | 0 | 8 | | 2 | 3,130 | 5,730 | 9,800 | 18,600 |
| 50 | +15 | 9 | | 1 | no sample | 20 | no sample | 13 |
| 50 | +15 | 9 | | 2 | no sample | 39 | no sample | 37 |
| 50 | +30 | 10 | | 1 | 299 | 2,960 | 18 | 28 |
| 50 | +30 | 10 | | 2 | 322 | 1,530 | 7.8 | 40 |
| Pooled variance for duplicates | | | | | 840,000 | 7,300,000 | 592,000,000 | 22,000,000 |
| F (Ratio of variances for not ground and ground) | | | | | | | 700 | 3.0 |

$$s_p^2 = \frac{1}{2k} \sum_{i=1}^k d_i^2$$

where d_i is the difference of k sets of duplicates (Ku 1969). Then we computed the ratio of the variances for the not-ground and ground sets of samples. For the <2-mm fraction, 2,4-DNT was detectable in all 15 duplicates for both the not-ground and ground samples, and the F ratio was 3.0. The critical value of $F_{(14,14)}$ is 2.48 ($P = 0.05$) (Miller and Miller 1984), so the machine grinding resulted in a significant increase in precision. The F ratio for the >2-mm fraction was highly significant ($F = 700$), but most of the variation was due to sample 3A, where the concentration estimates differed by a factor of 1800. Even excluding this one sample, machine grinding significantly improved precision. However, the reduction in subsampling variance by grinding the Bo-Whale sample is less than the reduction we find when unvegetated samples contaminated with high explosives, such as those collected from hand grenade ranges, were ground. For unvegetated samples contaminated with TNT, RDX, and HMX, the relative standard deviation for 12 replicates was less than 10% (Walsh et al. 2002).

To test if machine sample division would reduce the laboratory subsampling variance over that obtained by manual subsampling, we divided Bo-Whale samples 3A and 6 into 12 subsamples each using a rotary divider. For these samples, the relative standard deviations for the 2,4-DNT concentration estimates were 55% and 32%, respectively (Table 3). The pooled relative standard deviation for the 15 sets of duplicates of the ground <2-mm fractions of Bo-Whale samples 1–10 was 44% (Table 2), so machine division does not appear to improve subsampling precision for these samples. Future homogenization experiments will examine the effect of longer grind times on 2,4-DNT-contaminated soils.

To determine if we were able to collect field samples in a reproducible manner, we used the laboratory duplicates to compute the mean concentrations in the five sets of field duplicates for the >2-mm and <2-mm fractions with and without machine grinding. Again, using the ratio of the pooled variances (Table 4), we see that machine grinding significantly improved precision for both size fractions. The field replicates for the <2-mm machine-ground fractions were in relatively good agreement, considering the heterogeneity of the substrate we were sampling. However, methods to reduce the field sampling variance are needed.

We collected four sets of subsurface samples using an AMS soil core sampler to determine if propellant residues deposited from firing activities were migrating downward through the soil column. The locations of the subsurface samples were chosen based on the highest concentrations of 2,4-DNT detected using the field

Table 3. Subsampling heterogeneity in two machine ground samples that were split by a rotary divider.

| Replicate | 2,4-DNT Concentration ($\mu\text{g/kg}$) | |
|-----------|--|----------------------------------|
| | Bo-Whale Sample 6 (<2 mm) | Bo-Whale Sample 3A (<2 mm) |
| 1 | 7,400 | 810 |
| 2 | 4,900 | 1,860 |
| 3 | 6,800 | 860 |
| 4 | 3,900 | 2,900 |
| 5 | 4,200 | 3,530 |
| 6 | 8,000 | 1,700 |
| 7 | 3,500 | 2,500 |
| 8 | 7,000 | 1,150 |
| 9 | 6,097 | 4,200 |
| 10 | 6,000 | 1,900 |
| 11 | 2,650 | 920 |
| 12 | 4,300 | 1,600 |
| mean | 5,396 | 1,993 |
| min | 2,650 | 810 |
| max | 8,000 | 4,200 |
| median | 5,450 | 1,775 |
| RSD | 32% | 55% |

Table 4. Mean concentration estimates of the >2-mm and <2-mm fractions with and without machine grinding in field duplicate multi-increment samples at FP Bo-Whale.

| Distance from base plate (m) | Angle from centerline (degrees) | Sample ID | Field replicate | 2,4-DNT Conc. (µg/g) | | | |
|--|--|--------------|--------------------|----------------------|-----------|--------------|------------|
| | | | | Machine ground | | Not ground | |
| | | | | >2 mm | <2 mm | >2 mm | <2 mm |
| 3.5 | 0 | 1 | A | 1,230 | 7,000 | 7,990 | 3,360 |
| 3.5 | 0 | 1 | B | 349 | 3,520 | 1,770 | 1,310 |
| 7 | 0 | 2 | A | 136 | 2,200 | 718 | 2,750 |
| 7 | 0 | 2 | B | 948 | 3,341 | 1,680 | 3,740 |
| 14 | 0 | 3 | A | 549 | 1,240 | 217 | 368 |
| 14 | 0 | 3 | B | 154 | 1,700 | 61,550 | 7,020 |
| 21 | 0 | 4 | A | 467 | 1,200 | not detected | 540 |
| 21 | 0 | 4 | B | 2,170 | 1,900 | 290 | 196 |
| 28 | 0 | 5 | A | 3,660 | 23,400 | 11,200 | 22,750 |
| 28 | 0 | 5 | B | 13,100 | 26,600 | 10,300 | 9,410 |
| Pooled Variance for Duplicates | | | | 9,360,000 | 2,440,000 | 380,000,000 | 22,800,000 |
| F (Ratio of variances for not ground and ground) | | | | | | 41 | 9.4 |

GC analysis. Three sets were from FP Bo-Whale, and the fourth set was from FP Big Lake. The results in Table 5 show that the bulk of the residues were in the top 2 cm and that no analytes were detected below 5 cm deep.

Firing Points 2002

The firing point samples from 2001 showed that firing with 105-mm howitzers deposited 2,4-DNT on the surface soil in a heterogeneous manner resulting in parts-per-million residue concentrations and that the residue extended at least 50 m from the gun position. In 2002, we intensively sampled two howitzer firing positions, one vegetated and the other sparsely vegetated, shortly after the guns were used, and we repeated the sampling after 30 days. We must point out that the other guns at the firing points were positioned close enough so that some of the 2,4-DNT we detected may have been contributed by the firing of neighboring guns.

The range of 2,4-DNT concentrations at the sparsely vegetated gun position (FP Mark Gun 2) was <1–19,000 µg/kg shortly after firing in June and 2–32,000 µg/kg 30 days later in July (Table 6). At the vegetated gun position (FP Sally Gun 5) the range of 2,4-DNT concentrations was <1–5,800 µg/kg after

Table 5. Concentrations of propellant residues found in subsurface samples collected from FP Bo-Whale and Big Lake.

| | | | Concentration (µg/kg) | | |
|---|----------|-----|-----------------------|---------|-----|
| | Lab | Rep | 2,6-DNT | 2,4-DNT | NG |
| Bo-Whale FP Discrete Location 1 (within area BW4 composite sample) | | | | | |
| Surface | Field GC | | NA | 7,900 | NA |
| 0 to 2.5 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | 8.1 | <15 |
| 2.5 to 5 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 5 to 9 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 9 to 13 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| FP Bo-Whale Discrete Location 2 (within area BW4 composite sample) | | | | | |
| Surface | Field GC | | NA | 4,600 | <15 |
| 0 to 2.5 cm depth | Lab GC | A | 616 | 13,300 | 550 |
| | Lab GC | B | 588 | 11,300 | <15 |
| 2.5 to 5 cm depth | Lab GC | A | <1 | 19.6 | 250 |
| | Lab GC | B | <1 | 5.4 | <15 |
| 5 to 10 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 10 to 15 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| FP Bo-Whale Discrete Location 1.5 (within area BW4 composite sample) | | | | | |
| Surface | Lab GC | | 48.6 | 530 | <15 |
| 0 to 2 cm depth | Lab GC | A | 13.8 | 226 | <15 |
| | Lab GC | B | <1 | 8.3 | <15 |
| 2 to 4 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 4 to 11 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 11 to 15 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| FP Big Lake Discrete Location 10 (within area BL14 composite sample) | | | | | |
| Surface | Field GC | | NA | 9,100 | NA |
| Surface | Lab GC | | 345 | 6,790 | <15 |
| 1 to 4 cm depth | Lab GC | A | <1 | 4.0 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 4 to 8 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 8 to 15 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |
| 15 to 20 cm depth | Lab GC | A | <1 | <1 | <15 |
| | Lab GC | B | <1 | <1 | <15 |

Table 6. Concentrations of 2,4-DNT determined in composite surface soil samples collected around a 105-mm howitzer within one week (June 2002) and five weeks (July) of firing.

| FP Mark (sparsely vegetated) | | | | FP Sally (vegetated) | | | |
|-----------------------------------|---------------------------------|-----------------|--------|-----------------------------------|---------------------------------|-----------------|-------|
| Distance from firing platform (m) | Angle from centerline (degrees) | 2,4-DNT (µg/kg) | | Distance from firing platform (m) | Angle from centerline (degrees) | 2,4-DNT (µg/kg) | |
| | | June | July | | | June | July |
| 10 | 0 | 70 | 190 | 10 | 0 | 3,800 | 3,000 |
| 20 | 0 | 300 | 160 | 20 | 0 | 1,900 | 1,000 |
| 25 | 0 | 4,900 | 550 | 25 | 0 | 800 | 230 |
| 40 | 0 | 1,400 | 3,700 | 40 | 0 | 290 | 1600 |
| 50 | 0 | 250 | 150 | 50 | 0 | <d | 270 |
| 60 | 0 | 57 | 690 | 60 | 0 | <d | <d |
| 70 | 0 | 17 | 9.0 | 70 | 0 | <d | 260 |
| 80 | 0 | 1,400 | 110 | 10 | -30 | 2,200 | 7,400 |
| 90 | 0 | 120 | 1,100 | 25 | -30 | 1,100 | 2,700 |
| 100 | 0 | 300 | 1,200 | 50 | -30 | 70 | 60 |
| 10 | -30 | 120 | 120 | 10 | +30 | 810 | 2400 |
| 25 | -30 | 26 | 8 | 25 | +30 | 140 | 140 |
| 50 | -30 | 870 | 1,900 | 50 | +30 | 530 | 490 |
| 75 | -30 | 300 | 340 | 75 | +30 | <d | 64 |
| 100 | -30 | 4.0 | 36 | 100 | +30 | <d | <d |
| 10 | +30 | 110 | 250 | 10 | -60 | 5,800 | 4,400 |
| 25 | +30 | 1,800 | 1,800 | 25 | -60 | 450 | 1,500 |
| 50 | +30 | 2,000 | 2,300 | 50 | -60 | 240 | 63 |
| 75 | +30 | 2,300 | 1,400 | 10 | +60 | 86 | 750 |
| 95 | +30 | 3,600 | 3,300 | 25 | +60 | 670 | 810 |
| 10 | -60 | 240 | 950 | 50 | +60 | 190 | 100 |
| 25 | -60 | 1,400 | 2,900 | 75 | +60 | <d | 27 |
| 50 | -60 | 120 | 53 | 100 | +60 | <d | <d |
| 75 | -60 | 1,400 | 170 | 10 | -90 | 2,300 | 3,700 |
| 100 | -60 | 160 | 160 | 50 | -90 | 160 | 770 |
| 10 | +60 | 41 | 21 | 10 | +90 | 200 | 820 |
| 25 | +60 | 1,700 | 1,800 | 50 | +90 | <d | 140 |
| 50 | +60 | 170 | 440 | 10 | -120 | 620 | 1,400 |
| 75 | +60 | 1,500 | 1,800 | 50 | -120 | 1,400 | 900 |
| 100 | +60 | 19,000 | 32,000 | 10 | +120 | 32 | 210 |
| 10 | -90 | 120 | 100 | 50 | +120 | 35 | 94 |
| 50 | -90 | 42 | 140 | 10 | -150 | 230 | 160 |
| 10 | +90 | 72 | 68 | 50 | -150 | 180 | 750 |
| 50 | +90 | 67 | 270 | 10 | +150 | 220 | 360 |
| 10 | -120 | 50 | 4.0 | 50 | +150 | 26 | 62 |
| 36 | -120 | <d | 100 | 10 | 180 | 95 | 90 |
| 10 | +120 | 61 | 26 | 50 | 180 | 15 | <d |
| 50 | +120 | 1,000 | 940 | mean | | 660 | 990 |
| 10 | -150 | 7.0 | 2.0 | median | | 190 | 270 |
| 50 | -150 | <d | 3 | max | | 5,800 | 7,400 |
| 10 | +150 | 27 | 7.5 | | | | |
| 30 | +150 | 9.0 | 5.0 | | | | |
| 10 | 180 | 9.0 | 18 | | | | |
| 28 | 180 | 4.0 | 2.0 | | | | |
| mean | | 1,070 | 1,390 | | | | |
| median | | 121 | 165 | | | | |
| max | | 19,000 | 32,000 | | | | |

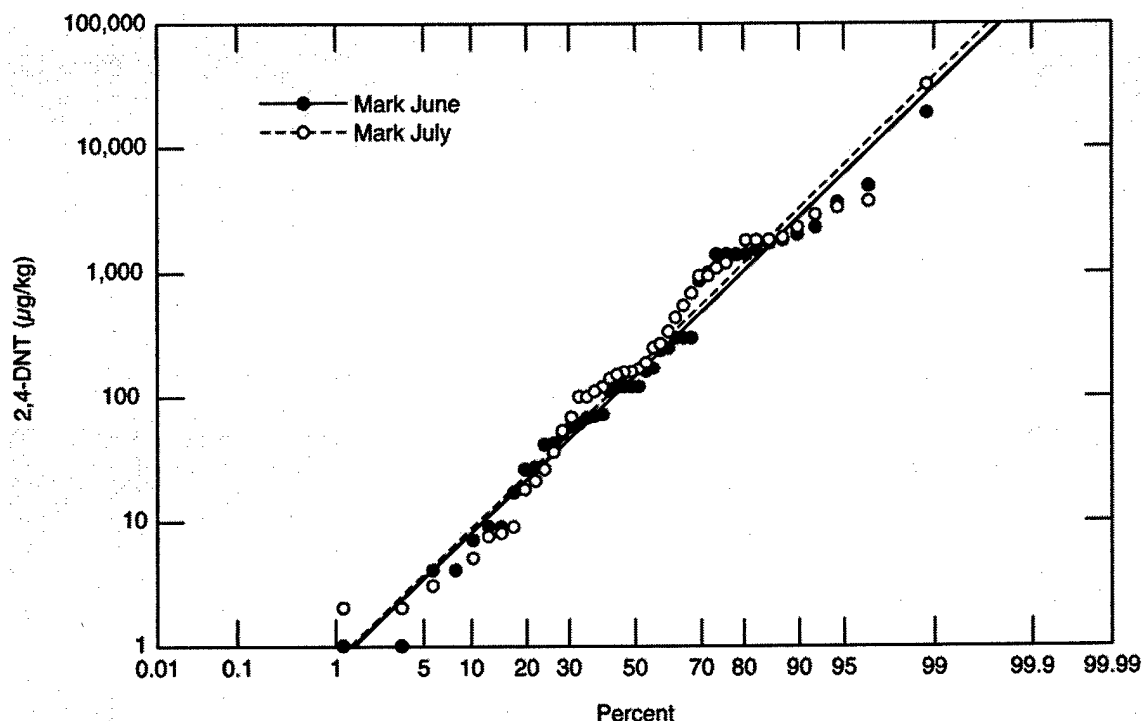


Figure 16. Probability plot of 2,4-DNT concentrations at FP Mark in June and July 2002. The data are log-normally distributed, and there was no significant change in 2,4-DNT concentration after 30 days of weathering.

firing and $<1\text{--}7,400\text{ }\mu\text{g/kg}$ 30 days later. The data were not normally distributed; when the data for FP Mark are displayed on a log probability plot (Fig. 16), the points fall approximately along straight lines. We used Wilcoxon Matched Pairs Test to compare the June and July concentrations estimates, and there was no significant difference for FP Mark. There was a significant difference between the June and July medians for FP Sally; the July median was greater than the June median, probably because we paid more attention to maintaining the sampling depth at only 1 cm for the July samples.

We did not detect 2,4-DNT in subsurface samples collected in July 2002 at FP Sally, the vegetated firing point. However, we could detect some 2,4-DNT in subsurface samples at FP Mark, which had sparse vegetation (Table 7). The organic matter in the vegetated soil would be expected to sorb any 2,4-DNT that dissolves in the surface moisture.

Samples from the other gun positions at FP Mark, Sally, Audrey, and Bo-Whale (Tables 8–11) in 2002 showed similar patterns for 2,4-DNT. With the exception of Bo-Whale gun positions one and two, 2,4-DNT was detectable at concentrations ranging from 10 to 8,800 $\mu\text{g/kg}$.

Table 7. Concentrations of 2,4-DNT determined in composite surface (0–1 cm) and subsurface (15–20 cm) soil samples collected near a 105-mm howitzer within five weeks (July 2002) after firing.

| Distance from firing platform (m) | Angle from centerline (degrees) | Depth | 2,4-DNT (µg/kg) | |
|-----------------------------------|---------------------------------|------------|-----------------|-------------|
| | | | Mark gun 2 | Sally gun 5 |
| 25 | 0 | Surface | 550 | 230 |
| | | Subsurface | 4.2 | <d |
| 50 | 0 | Surface | 150 | 270 |
| | | Subsurface | 17 | <d |
| 25 | –60 | Surface | 2,900 | 1,500 |
| | | Subsurface | 260 | <d |
| 50 | –60 | Surface | 53 | 63 |
| | | Subsurface | 59 | <d |
| 25 | +60 | Surface | 1,800 | 810 |
| | | Subsurface | 100 | <d |
| 50 | +60 | Surface | 440 | 100 |
| | | Subsurface | 250 | <d |

Table 8. Concentrations of 2,4-DNT detected at FP Mark in June 2002.

| Gun # | Distance from firing platform (m) | Angle from centerline (degrees) | 2,4-DNT (µg/kg) |
|-------|-----------------------------------|---------------------------------|-----------------|
| 1 | 25 | 0 | 1,250 |
| 1 | 50 | 0 | 1,000 |
| 1 | 25 | –60 | 410 |
| 1 | 50 | –60 | 200 |
| 1 | 25 | +60 | 2,750 |
| 1 | 50 | +60 | 2,200 |

Table 9. Concentrations of 2,4-DNT detected at FP Sally in June 2002.

| Gun # | Distance from firing platform (m) | Angle from centerline (degrees) | 2,4-DNT ($\mu\text{g/kg}$) |
|-------|-----------------------------------|---------------------------------|------------------------------|
| 1 | 25 | 0 | 62 |
| 1 | 50 | 0 | 110 |
| 1 | 25 | -60 | 255 |
| 1 | 50 | -60 | 740 |
| 1 | 25 | +60 | 520 |
| 1 | 50 | +60 | 4,800 |
| 2 | 25 | 0 | 225 |
| 2 | 50 | 0 | 8,800 |
| 2 | 25 | -60 | 765 |
| 2 | 50 | -60 | 3,900 |
| 2 | 50 | +60 | 1,500 |
| 2 | Shell case pile | | 5,800 |
| 3 | 25 | 0 | 3,300 |
| 3 | 50 | 0 | 480 |
| 3 | 25 | -60 | 480 |
| 3 | 50 | -60 | 165 |
| 3 | 25 | +60 | 520 |
| 3 | 50 | +60 | 3,200 |
| 4 | 25 | 0 | 170 |
| 4 | 50 | 0 | 10 |
| 4 | 25 | -60 | 830 |
| 4 | 50 | -60 | 2,400 |
| 4 | 25 | +60 | 1,500 |
| 4 | 50 | +60 | 790 |
| 6 | 25 | 0 | 815 |
| 6 | 50 | 0 | 490 |
| 6 | 25 | -60 | 66 |
| 6 | 50 | -60 | 110 |
| 6 | 25 | +60 | <d |
| 6 | 50 | +60 | 14 |

Table 10. Concentrations of 2,4-DNT detected at FP Audrey in June 2002.

| Gun # | Distance from firing platform (m) | Angle from centerline (degrees) | 2,4-DNT ($\mu\text{g/kg}$) |
|-------|-----------------------------------|---------------------------------|------------------------------|
| 1 | 25 | 0 | 590 |
| 1 | 50 | 0 | 1,200 |
| 1 | 25 | -60 | 77 |
| 1 | 50 | -60 | 170 |
| 1 | 25 | +60 | 330 |
| 1 | 50 | +60 | 46 |
| 2 | 25 | 0 | 570 |
| 2 | 40 | 0 | 1,700 |
| 2 | 25 | -60 | 2,100 |
| 2 | 50 | -60 | 870 |
| 2 | 25 | +60 | 70 |
| 2 | 44 | +60 | 180 |
| 3 | 25 | 0 | 1,100 |
| 3 | 50 | 0 | 80 |
| 3 | 25 | -60 and +60 | 110 |
| 3 | 50 | -60 and +60 | 390 |
| 4 | 25 | 0 | 1,700 |
| 4 | 50 | 0 | 670 |
| 4 | 25 | -60 and +60 | 360 |
| 4 | 50 | -60 and +60 | 570 |
| 5 | 20 | 0 | 710 |
| 5 | 25 | -60 and +60 | 230 |
| 5 | 50 | -60 and +60 | 90 |
| 6 | 25 | 0 | 1,900 |
| 6 | 25 | -60 | 6,800 |
| 6 | 50 | -60 | 240 |
| 6 | 25 | +60 | 10 |
| 6 | 35 | +60 | 110 |

Table 11. Concentrations of 2,4-DNT detected at FP Bo-Whale in June 2002.

| Gun # | Distance from firing platform (m) | Angle from centerline (degrees) | 2,4-DNT ($\mu\text{g/kg}$) |
|-------|-----------------------------------|---------------------------------|------------------------------|
| 1 | 25 | 0 | <d |
| 1 | 50 | 0 | <d |
| 1 | 25 | -60 | <d |
| 1 | 50 | -60 | <d |
| 1 | 25 | +60 | <d |
| 1 | 50 | +60 | <d |
| 2 | 25 | 0 | <d |
| 2 | 50 | 0 | 2,900 |
| 2 | 25 | -60 | 320 |
| 2 | 50 | -60 | 720 |
| 2 | 25 | +60 | <d |
| 2 | 50 | +60 | <d |
| 3 | 25 | 0 | 6,300 |
| 3 | 50 | 0 | 690 |
| 3 | 25 | -60 | 6,800 |
| 3 | 50 | -60 | 120 |
| 3 | 25 | +60 | 5,400 |
| 3 | 50 | +60 | 6,100 |
| 4 | 25 | 0 | 4,300 |
| 4 | 50 | 0 | <d |
| 4 | 25 | -60 | 570 |
| 4 | 50 | -60 | 1,500 |
| 4 | 25 | +60 | 1,000 |
| 4 | 50 | +60 | 620 |
| 5 | 25 | 0 | 470 |
| 5 | 50 | 0 | 1,400 |
| 5 | 25 | -60 | 700 |
| 5 | 50 | -60 | 400 |
| 5 | 25 | +60 | 830 |
| 5 | 50 | +60 | 1,100 |

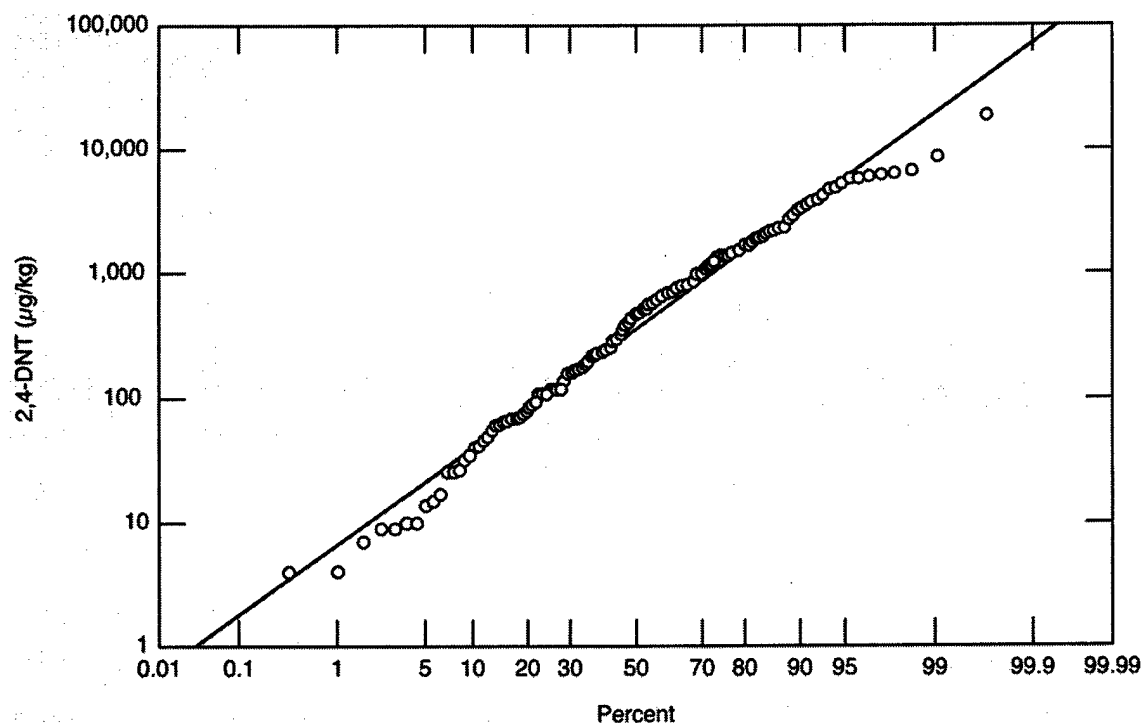


Figure 17. Probability plot of 2,4-DNT concentrations at FP Mark, Sally, Audrey, and Bo-Whale in June 2002. The data are log-normally distributed, and the median concentration was 480 µg/kg.

Pooling the data from FP Mark, Sally, Audrey, and Bo-Whale, we find 155 detections of 2,4-DNT out of the 175 samples collected in June 2002. The data were log-normally distributed (Fig. 17). The median concentration was 480 µg/kg.

The cartridge case for the 105-mm howitzer comes with a full complement of propellants arranged as seven individual bagged and numbered propelling charges (U.S. Army 1994). The distance the projectile is fired depends on the number of propelling charge increments. To fire at less than maximum range, excess propellant bags are removed. The previous practice was to burn these bags on the ground at the firing point. The current practice is to burn the excess propellant in pans at designated locations. The excess propellant for the training exercise in June 2002 was burned in a tray at Observation Point 7. The troops placed some soil in the tray so we could sample what would have been deposited on the soil surface if a tray had not been used. We also collected soil samples from the area downwind from the burn tray. The downwind side was to the southwest and was obvious from the dead leaves on the trees killed by the heat of the fire. Very high concentrations (2,300,000 µg/kg) of 2,4-DNT were detected in the soil from the burn tray 2 (Table 12). Downwind of the tray, concentrations were still high (120,000 µg/kg). We also detected 2,6-DNT in the burn samples,

with concentrations approximately 5% of the corresponding 2,4-DNT concentration.

The Lampkin Range firing point was used for direct fire of the 105-mm howitzers and for other munitions, including mortars. In the two composite samples we collected in July 2002, we found the same two analytes as those we detected in July 2000 (Walsh et al. 2001), namely 2,4-DNT and NG. The 2,4-DNT concentrations (260 and 370 $\mu\text{g/kg}$) were similar to those detected at the other firing points. NG was detected at 59,000 and 35,000 $\mu\text{g/kg}$.

Table 12. Concentrations of 2,4-DNT and 2,6-DNT in soil at Observation Point 7 where excess propellant was burned.

| | 2,4-DNT ($\mu\text{g/kg}$) | 2,6-DNT ($\mu\text{g/kg}$) |
|---------------------|---------------------------------|---------------------------------|
| Soil SW of Tray | 120,000 | 5,200 |
| Soil in Burn Tray 1 | 15,000 | 630 |
| Soil in Burn Tray 2 | 2,300,000 | 130,000 |

Collection of Propellant Residue from a Snow-covered Firing Point

We detected 2,4-DNT and 2,6-DNT in each of the surface snow samples (Table 13) we collected immediately after the winter firing of 105-mm projectiles (Fig. 13). We computed the equivalent soil concentrations based on the mass of residue deposited in each 1- m^2 sample area. Assuming that the residues reside in the top 1 cm of soil and that the bulk density of the soil is 1.5 g/cm^3 , then the mass of soil containing residue in each 1- m^2 area would be 15 kg. For 2,4-DNT the range of soil concentrations in front of the howitzer would have been 22–1,900 $\mu\text{g/kg}$, with a median of 430 $\mu\text{g/kg}$, which is very similar to the median soil concentration for FP Mark, Sally, Audrey, and Bo-Whale (480 $\mu\text{g/kg}$). The variability of concentrations in neighboring snow samples is also similar to the variability in the soil samples from Donnelly Training Area.

Table 13. 2,4-DNT and 2,6-DNT concentrations detected on snow following the firing of 105-mm howitzers and the equivalent[†] soil concentration.

| Sample ID | Distance from firing platform (m) | Angle from centerline (degrees) | 2,4-DNT | | 2,6-DNT | |
|-----------|-----------------------------------|---------------------------------|--|--|--|--|
| | | | Conc. found on snow ($\mu\text{g}/\text{m}^2$) | Equivalent [†] soil conc. ($\mu\text{g}/\text{kg}$) | Conc. found on snow ($\mu\text{g}/\text{m}^2$) | Equivalent [†] soil conc. ($\mu\text{g}/\text{kg}$) |
| 1 | 4 | +40 | 16,500 | 1,100 | 1,120 | 75 |
| 4 | 5 | -10 | 15,400 | 1,027 | 1,060 | 71 |
| 2 | 6 | +40 | 9,250 | 617 | 544 | 36 |
| 7 | 6 | -40 | 28,200 | 1,880 | 1,510 | 101 |
| 3 | 8 | +10 | 920 | 61 | 39 | 3 |
| 6 | 8 | +30 | 2,770 | 185 | 158 | 11 |
| 15 | 9 | +15 | 9,980 | 665 | 674 | 45 |
| 16 | 12 | -20 | 13,800 | 920 | 882 | 59 |
| 8 | 13 | +10 | 3,660 | 244 | 236 | 16 |
| 10 | 14 | +30 | 1,060 | 71 | 69 | 5 |
| 17 | 15 | -50 | 11,200 | 747 | 418 | 28 |
| 12 | 23 | +15 | 494 | 33 | 27 | 2 |
| 13 | 23 | +10 | 336 | 22 | 19 | 1 |
| 14 | 25 | -10 | 744 | 50 | 29 | 2 |
| 5 | Gun 3 Breach | | 305 | 20 | 14 | 1 |
| 9 | Gun 2 Breach | | 162 | 11 | 12 | 1 |
| 11 | Gun 4 Breach | | 1,430 | 95 | 55 | 4 |

[†]Assuming that 1-m² of soil with a bulk density of 1.5 g/cm³ is sampled to a depth of 1 cm, the mass of soil would be 15 kg.

6 DISCUSSION

Explosives Residues on Impact Areas

Two of the impact areas that we sampled (Georgia Island and Washington Range West) did not have detectable concentrations of explosives. Georgia Island has not been used for a number of years, and Washington Range West is really a buffer zone for the Washington Impact Area. On Delta Creek, the spatial distribution of explosives residues was similar to what has been observed on other active impact areas. Explosives residues, if detectable at all, are at very low concentrations (parts per billion) over most of the ranges. In contrast, localized areas where ordnance has failed to completely detonate may have solid explosives on the soil surface, and the underlying soil can have high parts-per-million concentrations. Targets, where ordnance detonations are concentrated, can also have detectable concentrations of explosives. On Delta Creek, we found localized high concentrations of TNT, the high-explosive filler of 500-lb bombs. We also found RDX, which could have come from a variety of ordnance items (Table 1), including C4, which is used to detonate unexploded ordnance. NG was also detected in soil under rocket motors. At Delta Creek, explosives residues from range scrap and partially detonated ordnance can move to the surface water by erosion of the floodplain terrace (Fig. 2b).

Propellant Residues at Firing Points

Unlike impact areas, where ordnance residues are for the most part undetectable, each of the howitzer firing points that we have sampled at the Donnelly Training Area and elsewhere have detectable concentrations of 2,4-DNT. The data were log-normally distributed, with median concentrations around 500 µg/kg.

The Agency for Toxic Substances and Disease Registry published a toxicological profile for 2,4-DNT and 2,6-DNT in December 1998 that summarizes information on the adverse health effects and numerous regulations associated with these compounds (Science International Inc. 1998). Munitions workers with chronic DNT exposure had a variety of health problems affecting the circulatory and nervous systems. Both 2,4- and 2,6-DNT caused liver cancer in laboratory animals, and the International Agency for Research on Cancer (IARC) has designated that these chemicals are probable human carcinogens, based on animal data (Group B2) (Science International Inc. 1998). The EPA-derived oral reference doses (RfDs), which are not applicable to cancer risk, are 0.002 mg/kg/day for 2,4-DNT and 0.001 mg/kg/day for 2,6-DNT. Based on these RfDs, the Drinking Water Equivalent Levels are 0.1 and 0.04 mg/L for 2,4-DNT and 2,6-DNT, respectively. Lifetime drinking water advisory values are not listed due to the cancer risk.

The EPA Region III Risk-Based Concentration Table gives soil screening levels for the protection of groundwater based on non-carcinogenic effects (U.S. EPA 2003). For 2,4-DNT and 2,6-DNT (an impurity in military-grade TNT and 2,4-DNT), these values are 29 and 12 $\mu\text{g}/\text{kg}$ for 2,4-DNT and 2,6-DNT, respectively, if the dilution attenuation factor is one, and 570 and 250 $\mu\text{g}/\text{kg}$ for 2,4-DNT and 2,6-DNT, respectively, if the dilution attenuation factor is 20.

In the last few years, states, including Alaska, have issued soil cleanup levels for 2,4-DNT, 2,6-DNT, and several other chemicals. The State of Alaska (Alaska Department of Environmental Conservation 2002) has three sets of soil cleanup standards that are based on climate zones: Arctic (continuous permafrost); Under 40 Inch Zone [less than 40 inches (102 cm) of annual precipitation]; and Over 40 Inch Zone [greater than 40 inches (102 cm) of annual precipitation]. The Big Delta National Weather Service Station receives an average of 12 inches (30 cm) of precipitation a year, so the Donnelly Training Area is in the Under 40 Inch Zone. Alaska Department of Environmental Conservation Title 18 Alaska Administrative Code Chapter 75 lists 2,4-DNT and 2,6-DNT as carcinogenic chemicals. As a result, the soil cleanup standards are extremely low for the protection of groundwater: 5 $\mu\text{g}/\text{kg}$ for 2,4-DNT and 4.4 $\mu\text{g}/\text{kg}$ for 2,6-DNT. The equations and input parameters used to derive these values are described in *Guidance on Cleanup Levels Equations and Input Parameters* (Alaska Department of Environmental Conservation 1999).

Most of the samples at firing points Sally, Mark, Audrey, and Bo-Whale had concentrations of 2,4-DNT that exceeded the Alaska soil cleanup levels by a wide margin. Alternative cleanup levels that are based on site-specific soil data and an approved fate and transport model may be approved if the alternative cleanup levels are "protective of human health, safety, and welfare and the environment" (Alaska Department of Environmental Conservation 2002). The alternative levels must not exceed the ingestion-based levels, which are 12,000 $\mu\text{g}/\text{kg}$ for 2,4-DNT and 2,6-DNT. Most of the samples from the firing points were less than 12,000 $\mu\text{g}/\text{kg}$, but the propellant burn area far exceeded this level. The subsurface samples we collected indicated that downward migration of these contaminants was minimal, but prudent placement of firing points and especially propellant burn locations is desirable because of the low screening levels given for protection of groundwater.

The compound 2,4-DNT biotransforms in the environment and ultimately mineralizes through reductive and/or oxidative pathways. The persistence of 2,4-DNT associated with unburned propellant compositions is unknown, but it is probably enhanced by 2,4-DNT's residence within a nitrocellulose matrix. Nitrocellulose is insoluble in water and could only migrate to surface water by bulk movement of solids by water or wind.

7 CONCLUSIONS

We sampled some impact areas of the Donnelly Training Area using authoritative sampling, when possible, to try to detect explosives residues in surface soils. We did not detect explosives residues on Georgia Island and Washington Range West. We did detect NG, a propellant residue, in one discrete sample collected under a 40-mm cartridge case on Georgia Island. The target array downstream of the Delta Creek Impact Area appeared to be more heavily used than the previous two areas, and we found explosives residues in all of the samples collected around craters, targets, and ordnance debris. This impact area had been used by the Air Force for training with 500- and 2000-lb bombs, and partial detonations of these bombs created localized areas containing high concentrations of TNT. RDX was detected in several samples; the two highest RDX concentrations were associated with targets. We did not detect TNT, RDX, or other high-explosives residues in composite soil samples collected upstream and downstream from the target array. We did detect NG in discrete samples downstream from the target array; these discrete samples were collected under pieces of rockets. Explosives residues were detectable in each of the soils samples collected from a MOUT/CALFEX site. Specifically, NG was associated with 40-mm grenade training, and low concentrations of TNT, RDX, and 2,4-DNT were associated with explosive ordnance disposal craters.

Soils from recently used firing points have parts-per-million concentrations of NG and 2,4-DNT. These residues are most likely associated with partially burned propellant. The 2,4-DNT is found on the surface of vegetated firing points, and we could not detect any decrease in 2,4-DNT concentrations after 30 days of weathering at either vegetated or sparsely vegetated firing points. Results from replicate field and laboratory samples for 2,4-DNT indicate that sampling error is high; research to improve field and laboratory sampling is ongoing. The highest concentrations of 2,4-DNT were in soils where excess propellant is burned. Fixed firing points and propellant burn areas should be located away from groundwater recharge areas.

Both 2,4-DNT and 2,6-DNT are listed as hazardous substances by the State of Alaska, and very low soil cleanup levels for the protection of groundwater are given for these potentially carcinogenic compounds. Future work will focus on sample collection methods appropriate to obtain average concentrations over a firing point to provide data for possible risk assessment activities.

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Appendix A (cont.).

| Sample Type | Collector | Uniq. ID | Field Notes | Date Collected | Area | East (m) | North (m) | Elevation (m) | Field ID | US Top | US 144X | RDX | TNB | DNB | TEPRY | TNT | 44-DNT | 24-DNT | 28-DNT | 40-DNT | 48-DNT | 54-DNT | 60-DNT | 66-DNT | 72-DNT | 78-DNT | 84-DNT | 90-DNT | 96-DNT | 102-DNT | 108-DNT | 114-DNT | 120-DNT | 126-DNT | 132-DNT | 138-DNT | 144-DNT | 150-DNT | 156-DNT | 162-DNT | 168-DNT | 174-DNT | 180-DNT | 186-DNT | 192-DNT | 198-DNT | 204-DNT | 210-DNT | 216-DNT | 222-DNT | 228-DNT | 234-DNT | 240-DNT | 246-DNT | 252-DNT | 258-DNT | 264-DNT | 270-DNT | 276-DNT | 282-DNT | 288-DNT | 294-DNT | 300-DNT | 306-DNT | 312-DNT | 318-DNT | 324-DNT | 330-DNT | 336-DNT | 342-DNT | 348-DNT | 354-DNT | 360-DNT | 366-DNT | 372-DNT | 378-DNT | 384-DNT | 390-DNT | 396-DNT | 402-DNT | 408-DNT | 414-DNT | 420-DNT | 426-DNT | 432-DNT | 438-DNT | 444-DNT | 450-DNT | 456-DNT | 462-DNT | 468-DNT | 474-DNT | 480-DNT | 486-DNT | 492-DNT | 498-DNT | 504-DNT | 510-DNT | 516-DNT | 522-DNT | 528-DNT | 534-DNT | 540-DNT | 546-DNT | 552-DNT | 558-DNT | 564-DNT | 570-DNT | 576-DNT | 582-DNT | 588-DNT | 594-DNT | 600-DNT | 606-DNT | 612-DNT | 618-DNT | 624-DNT | 630-DNT | 636-DNT | 642-DNT | 648-DNT | 654-DNT | 660-DNT | 666-DNT | 672-DNT | 678-DNT | 684-DNT | 690-DNT | 696-DNT | 702-DNT | 708-DNT | 714-DNT | 720-DNT | 726-DNT | 732-DNT | 738-DNT | 744-DNT | 750-DNT | 756-DNT | 762-DNT | 768-DNT | 774-DNT | 780-DNT | 786-DNT | 792-DNT | 798-DNT | 804-DNT | 810-DNT | 816-DNT | 822-DNT | 828-DNT | 834-DNT | 840-DNT | 846-DNT | 852-DNT | 858-DNT | 864-DNT | 870-DNT | 876-DNT | 882-DNT | 888-DNT | 894-DNT | 900-DNT | 906-DNT | 912-DNT | 918-DNT | 924-DNT | 930-DNT | 936-DNT | 942-DNT | 948-DNT | 954-DNT | 960-DNT | 966-DNT | 972-DNT | 978-DNT | 984-DNT | 990-DNT | 996-DNT | 1002-DNT | 1008-DNT | 1014-DNT | 1020-DNT | 1026-DNT | 1032-DNT | 1038-DNT | 1044-DNT | 1050-DNT | 1056-DNT | 1062-DNT | 1068-DNT | 1074-DNT | 1080-DNT | 1086-DNT | 1092-DNT | 1098-DNT | 1104-DNT | 1110-DNT | 1116-DNT | 1122-DNT | 1128-DNT | 1134-DNT | 1140-DNT | 1146-DNT | 1152-DNT | 1158-DNT | 1164-DNT | 1170-DNT | 1176-DNT | 1182-DNT | 1188-DNT | 1194-DNT | 1200-DNT | 1206-DNT | 1212-DNT | 1218-DNT | 1224-DNT | 1230-DNT | 1236-DNT | 1242-DNT | 1248-DNT | 1254-DNT | 1260-DNT | 1266-DNT | 1272-DNT | 1278-DNT | 1284-DNT | 1290-DNT | 1296-DNT | 1302-DNT | 1308-DNT | 1314-DNT | 1320-DNT | 1326-DNT | 1332-DNT | 1338-DNT | 1344-DNT | 1350-DNT | 1356-DNT | 1362-DNT | 1368-DNT | 1374-DNT | 1380-DNT | 1386-DNT | 1392-DNT | 1398-DNT | 1404-DNT | 1410-DNT | 1416-DNT | 1422-DNT | 1428-DNT | 1434-DNT | 1440-DNT | 1446-DNT | 1452-DNT | 1458-DNT | 1464-DNT | 1470-DNT | 1476-DNT | 1482-DNT | 1488-DNT | 1494-DNT | 1500-DNT | 1506-DNT | 1512-DNT | 1518-DNT | 1524-DNT | 1530-DNT | 1536-DNT | 1542-DNT | 1548-DNT | 1554-DNT | 1560-DNT | 1566-DNT | 1572-DNT | 1578-DNT | 1584-DNT | 1590-DNT | 1596-DNT | 1602-DNT | 1608-DNT | 1614-DNT | 1620-DNT | 1626-DNT | 1632-DNT | 1638-DNT | 1644-DNT | 1650-DNT | 1656-DNT | 1662-DNT | 1668-DNT | 1674-DNT | 1680-DNT | 1686-DNT | 1692-DNT | 1698-DNT | 1704-DNT | 1710-DNT | 1716-DNT | 1722-DNT | 1728-DNT | 1734-DNT | 1740-DNT | 1746-DNT | 1752-DNT | 1758-DNT | 1764-DNT | 1770-DNT | 1776-DNT | 1782-DNT | 1788-DNT | 1794-DNT | 1800-DNT | 1806-DNT | 1812-DNT | 1818-DNT | 1824-DNT | 1830-DNT | 1836-DNT | 1842-DNT | 1848-DNT | 1854-DNT | 1860-DNT | 1866-DNT | 1872-DNT | 1878-DNT | 1884-DNT | 1890-DNT | 1896-DNT | 1902-DNT | 1908-DNT | 1914-DNT | 1920-DNT | 1926-DNT | 1932-DNT | 1938-DNT | 1944-DNT | 1950-DNT | 1956-DNT | 1962-DNT | 1968-DNT | 1974-DNT | 1980-DNT | 1986-DNT | 1992-DNT | 1998-DNT | 2004-DNT | 2010-DNT | 2016-DNT | 2022-DNT | 2028-DNT | 2034-DNT | 2040-DNT | 2046-DNT | 2052-DNT | 2058-DNT | 2064-DNT | 2070-DNT | 2076-DNT | 2082-DNT | 2088-DNT | 2094-DNT | 2100-DNT | 2106-DNT | 2112-DNT | 2118-DNT | 2124-DNT | 2130-DNT | 2136-DNT | 2142-DNT | 2148-DNT | 2154-DNT | 2160-DNT | 2166-DNT | 2172-DNT | 2178-DNT | 2184-DNT | 2190-DNT | 2196-DNT | 2202-DNT | 2208-DNT | 2214-DNT | 2220-DNT | 2226-DNT | 2232-DNT | 2238-DNT | 2244-DNT | 2250-DNT | 2256-DNT | 2262-DNT | 2268-DNT | 2274-DNT | 2280-DNT | 2286-DNT | 2292-DNT | 2298-DNT | 2304-DNT | 2310-DNT | 2316-DNT | 2322-DNT | 2328-DNT | 2334-DNT | 2340-DNT | 2346-DNT | 2352-DNT | 2358-DNT | 2364-DNT | 2370-DNT | 2376-DNT | 2382-DNT | 2388-DNT | 2394-DNT | 2400-DNT | 2406-DNT | 2412-DNT | 2418-DNT | 2424-DNT | 2430-DNT | 2436-DNT | 2442-DNT | 2448-DNT | 2454-DNT | 2460-DNT | 2466-DNT | 2472-DNT | 2478-DNT | 2484-DNT | 2490-DNT | 2496-DNT | 2502-DNT | 2508-DNT | 2514-DNT | 2520-DNT | 2526-DNT | 2532-DNT | 2538-DNT | 2544-DNT | 2550-DNT | 2556-DNT | 2562-DNT | 2568-DNT | 2574-DNT | 2580-DNT | 2586-DNT | 2592-DNT | 2598-DNT | 2604-DNT | 2610-DNT | 2616-DNT | 2622-DNT | 2628-DNT | 2634-DNT | 2640-DNT | 2646-DNT | 2652-DNT | 2658-DNT | 2664-DNT | 2670-DNT | 2676-DNT | 2682-DNT | 2688-DNT | 2694-DNT | 2700-DNT | 2706-DNT | 2712-DNT | 2718-DNT | 2724-DNT | 2730-DNT | 2736-DNT | 2742-DNT | 2748-DNT | 2754-DNT | 2760-DNT | 2766-DNT | 2772-DNT | 2778-DNT | 2784-DNT | 2790-DNT | 2796-DNT | 2802-DNT | 2808-DNT | 2814-DNT | 2820-DNT | 2826-DNT | 2832-DNT | 2838-DNT | 2844-DNT | 2850-DNT | 2856-DNT | 2862-DNT | 2868-DNT | 2874-DNT | 2880-DNT | 2886-DNT | 2892-DNT | 2898-DNT | 2904-DNT | 2910-DNT | 2916-DNT | 2922-DNT | 2928-DNT | 2934-DNT | 2940-DNT | 2946-DNT | 2952-DNT | 2958-DNT | 2964-DNT | 2970-DNT | 2976-DNT | 2982-DNT | 2988-DNT | 2994-DNT | 3000-DNT |
|-------------|-----------|----------|-------------|----------------|------|----------|-----------|---------------|----------|--------|---------|-----|-----|-----|-------|-----|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|-----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SE = not confirmed
NA = not analysed for this compound
ND = not detected
NS = not searched

Appendix A (cont.).

| Sample Type | Collector | Unique ID | Field Notes | Date Collected | Area | East (m) | North (m) | Elevation (m) | Lab Notes or ID | Lab Rep Units | LR | ROX | THS | DHS | TEHYL | TNT | 4L-DNT | 2A-DNT | 2B-DNT | 2L-DNT | NB | 2AT | 3AT | 4AT | NO | 3,5-DNA | |
|-------------|--------------|-----------|-----------------------------------|----------------|-----------|-----------|-------------|---------------|------------------------|---------------|----|------|-----|-------|-------|-------|--------|--------|--------|--------|-------|-------|-------|-------|-------|---------|-------|
| Composite | TJ-KM-MC-MEW | BW3-A | 14 m from Base Plate | 8/2/2001 | Row White | 555,822.3 | 7,082,261.2 | 503.0 | HPLC and GC, CHREL Lab | BW3-A | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 26.6 | 732 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,822.3 | 7,082,261.2 | 503.0 | HPLC and GC, CHREL Lab | BW3-B | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 46 | 1400 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW3-B | 14 m from Base Plate | 8/2/2001 | Row White | 555,822.3 | 7,082,261.2 | 503.0 | HPLC and GC, CHREL Lab | BW3-B | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 77.9 | 2010 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,818.2 | 7,082,267.4 | 502.8 | HPLC and GC, CHREL Lab | BW4-A | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 35.7 | 1280 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW4-A | 21 m from Base Plate | 8/2/2001 | Row White | 555,818.2 | 7,082,267.4 | 502.8 | HPLC and GC, CHREL Lab | BW4-A | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 31 | 1120 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,819.2 | 7,082,267.4 | 502.8 | HPLC and GC, CHREL Lab | BW4-B | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 51.9 | 1520 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW4-B | 21 m from Base Plate | 8/2/2001 | Row White | 555,818.2 | 7,082,267.4 | 502.8 | HPLC and GC, CHREL Lab | BW4-B | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 90.1 | 2300 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,816.2 | 7,082,273.7 | 502.8 | HPLC and GC, CHREL Lab | BW5-A | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 763 | 6680 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW5-A | 28 m from Base Plate | 8/2/2001 | Row White | 555,816.2 | 7,082,273.7 | 502.8 | HPLC and GC, CHREL Lab | BW5-A | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 1300 | 28900 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,816.2 | 7,082,273.7 | 502.8 | HPLC and GC, CHREL Lab | BW5-B | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 1180 | 24000 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW5-B | 28 m from Base Plate | 8/2/2001 | Row White | 555,816.2 | 7,082,273.7 | 502.8 | HPLC and GC, CHREL Lab | BW5-B | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 1330 | 29100 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,787.2 | 7,082,278.3 | 499.3 | HPLC and GC, CHREL Lab | BW6 | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 133 | 4020 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW6 | 50 m from Base Plate (30 degrees) | 8/2/2001 | Row White | 555,787.2 | 7,082,278.3 | 499.3 | HPLC and GC, CHREL Lab | BW6 | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 126 | 2640 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,787.2 | 7,082,278.3 | 499.3 | HPLC and GC, CHREL Lab | BW7 | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 365 | 8320 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW7 | 50 m from Base Plate (15 degrees) | 8/2/2001 | Row White | 555,796.6 | 7,082,280.0 | 499.7 | HPLC and GC, CHREL Lab | BW7 | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 235 | 5880 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,805.6 | 7,082,283.9 | 501.1 | HPLC and GC, CHREL Lab | BW8 | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 219 | 8790 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW8 | 50 m from Base Plate (0 degrees) | 8/2/2001 | Row White | 555,805.6 | 7,082,283.9 | 501.1 | HPLC and GC, CHREL Lab | BW8 | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 268 | 5780 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,818.1 | 7,082,285.5 | 499.9 | HPLC and GC, CHREL Lab | BW9 | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 0 | 20 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW9 | 50 m from Base Plate (15 degrees) | 8/2/2001 | Row White | 555,818.1 | 7,082,285.5 | 499.9 | HPLC and GC, CHREL Lab | BW9 | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 0 | 39 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,818.1 | 7,082,285.5 | 499.9 | HPLC and GC, CHREL Lab | BW10 | 1 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 120 | 2860 | NA | NA | NA | <15 |
| Composite | TJ-KM-MC-MEW | BW10 | 50 m from Base Plate (30 degrees) | 8/2/2001 | Row White | 555,830.3 | 7,082,292.2 | 502.6 | HPLC and GC, CHREL Lab | BW10 | 2 | µg/g | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | 64 | 1530 | NA | NA | NA | <15 | <5 |
| | | | | 8/2/2001 | Row White | 555,830.3 | 7,082,292.2 | 502.6 | HPLC and GC, CHREL Lab | BW11-A | 1 | µg/g | NR | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 |
| Composite | TJ-KM-MC-MEW | BW11 | 3.5 m from Base Plate | 8/2/2001 | Row White | 555,981.6 | 7,082,212.3 | 503.1 | 100417 | BW11-A | 1 | µg/g | NR | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | |
| | | | | 8/2/2001 | Row White | 555,981.6 | 7,082,212.3 | 503.1 | 100418 | BW11-B | 1 | µg/g | NR | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 | <25.3 |
| Composite | TJ-KM-MC-MEW | BW12 | 7 m from Base Plate | 8/2/2001 | Row White | 555,988.0 | 7,082,215.3 | 503.9 | 100419 | BW12-A | 1 | µg/g | NR | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | <23.4 | |
| | | | | 8/2/2001 | Row White | 555,988.0 | 7,082,215.3 | 503.9 | 100420 | BW12-B | 1 | µg/g | NR | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 | <23.5 |
| Composite | TJ-KM-MC-MEW | BW13 | 14 m from Base Plate | 8/2/2001 | Row White | 555,978.7 | 7,082,221.0 | 503.9 | 100421 | BW13-A | 1 | µg/g | NR | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | <24.9 | |
| | | | | 8/2/2001 | Row White | 555,978.7 | 7,082,221.0 | 503.9 | 100422 | BW13-B | 1 | µg/g | NR | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 | <25.1 |
| Composite | TJ-KM-MC-MEW | BW14 | 21 m from Base Plate | 8/2/2001 | Row White | 555,872.8 | 7,082,227.0 | 504.4 | 100423 | BW14-A | 1 | µg/g | NR | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | <25.0 | |
| | | | | 8/2/2001 | Row White | 555,872.8 | 7,082,227.0 | 504.4 | 100424 | BW14-B | 1 | µg/g | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 | <25 |
| Composite | TJ-KM-MC-MEW | BW15 | 28 m from Base Plate | 8/2/2001 | Row White | 555,968.6 | 7,082,232.7 | 504.5 | 100425 | BW15-A | 1 | µg/g | NR | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | |
| | | | | 8/2/2001 | Row White | 555,968.6 | 7,082,232.7 | 504.5 | 100425 | BW15-B | 1 | µg/g | NR | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 | <24.6 |

= not confirmed
NA = not analyzed for this compound
ND = not detected
NR = not reported

Appendix A (cont.).

[illegible]

† = not confirmed
 ‡A = not analysed for this compound
 †D = not detected
 †S = not reported

Appendix A (cont.). Analytical results from 2001.

[illegible]

† = not confirmed
 ‡A = not analyzed for this compound
 ND = not detected
 ‡B = not resolved

Appendix A (cont.).

| Sample Type | Collector | Unique ID | Field Notes | Date Collected | Area | East (m) | North (m) | Elevation (m) | Lab Notes or ID | Field ID | Rep | Units | HMX | RDX | TNB | DNB | TEOTYL | TNT | 4A-DNT | 2A-DNT | 2,6-DNT | 2,4-DNT | NB | 2,NT | 3,NT | 4,NT | NG | 3,5-DNA |
|----------------------------------|-----------|-----------|---|----------------|-------------|-----------|-------------|---------------|-----------------|----------------------|-----|-------|-----|-----|-----|-----|--------|-----|--------|--------|---------|---------|----|------|------|------|-----|---------|
| Discrete/ Subdiscrete/ Discrete/ | JAS-SH | BL10 | DISCRET | 9/4/2001 | Bog Lake PP | 554,892.2 | 7,080,781.1 | 491.5 | CORREL Lab GC | Subdiscrete Discrete | B | µg/kg | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | NA | NA | NA | NA | <15 | <5 |
| Discrete/ | JAS-SH | BL10 | DISCRET | 9/4/2001 | Bog Lake PP | 554,892.2 | 7,080,781.1 | 491.5 | CORREL Lab GC | BL10 within BL14 | A | µg/kg | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | NA | NA | NA | NA | <15 | <5 |
| Discrete/ | JAS-SH | BL10 | DISCRET | 9/4/2001 | Bog Lake PP | 554,892.2 | 7,080,781.1 | 491.5 | CORREL Lab GC | BL10 within BL14 | B | µg/kg | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | <5 | NA | NA | NA | NA | <15 | <5 |
| Discrete | MW | Discrete | Surface (Within area BL14 composite sample) | | | | | | CORREL Field GC | BL7 Discrete | | µg/kg | NA | NA | NA | NA | NA | NA | ND | NA | NA | NA | NA | NA | NA | NA | NA | NA |

< = not confirmed
 NA = not analyzed for this compound
 ND = not detected
 NR = not reported